## Assembling Genomes

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| (Translating the cloning jargon) |  |  |
| :---: | :---: | :---: |
| CLONE LIBRARIES USED FOR GENOME MAPPING AND SEQUENCING |  |  |
| Vector | Human-DNA insert size range | Number of clones required to cover the human genome |
| Yeast artificial chromosome (YAC) | $100-2,000 \mathrm{~kb}$ | 3,000 (1,000 kb) |
| Bacterial artificial chromosome (BAC) | 80-350 kb | 20,000 (150 kb) |
| Cosmid | 30-45 kb | 75,000 (40 kb) |
| Plasmid | 3-10 kb | 600,000 (5 kb) |
| M13 phage | 1 kb | 3,000,000 (1 kb) |

## Thinking about the basic shotgun concept

- Start with a very large set of random sequencing reads
- How might we match up the overlapping sequences?
- How can we assemble the overlapping reads together in order to derive the genome?


## Thinking about the basic shotgun concept

- At a high level, the first genomes were sequenced by comparing pairs of reads to find overlapping reads
- Then, building a graph (i.e., a network) to represent those relationships
- The genome sequence is a "walk" across that graph


## The "Overlap-Layout-Consensus" method

Overlap: Compare all pairs of reads
(allow some low level of mismatches)
Layout: Construct a graph describing the overlaps


Simplify the graph read
Find the simplest path through the graph
Consensus: Reconcile errors among reads along that path to find the consensus sequence



Overlap graph


## Simplifying an overlap graph



1. Remove all contained nodes \& edges going to them

## Simplifying an overlap graph


2. Transitive edge removal:

Given $A-B-C$ and $A-C$, remove $A-C$

## Simplifying an overlap graph


3. If un-branched, calculate consensus sequence If branched, assemble un-branched bits and then decide how they fit together

## Simplifying an overlap graph


"contig" (assembled contiguous sequence)

## This basic strategy was used for most of the early genomes. <br> Also useful: "mate pairs"



Contigs can be ordered using these paired reads


## GigAssembler (used to assemble the public human genome project sequence)



Jim Kent David Haussler

## Whole genome Assembly: big picture



## GigAssembler - Preprocessing

1. Decontaminating \& Repeat Masking.
2. Aligning of mRNAs, ESTs, BAC ends \& paired reads against initial sequence contigs.

- psLayout $\rightarrow$ BLAT

3. Creating an input directory (folder) structure.



# GigAssembler: Build merged sequence contigs ("rafts") 



Figure 1 Two sequences overlapping end to end. The sequences are represented as dashes. The aligning regions are joined by vertical bars. End-to-end overlap is an extremely strong indication that two sequences should be joined into a contig.

## Sequencing quality (Phred Score)



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$$
\begin{aligned}
& Q=-10 \log _{10} P \longleftarrow \begin{array}{l}
\text { Base-calling } \\
\text { or } \\
\text { Error } \\
\text { Probability }
\end{array} \\
& P=10^{\frac{-Q}{10}}
\end{aligned}
$$

Phred quality scores are logarithmically linked to error probabilities
Phred Quality Score Probability of incorrect base call Base call accuracy

| 10 | 1 in 10 | $90 \%$ |
| :--- | :--- | :--- |
| 20 | 1 in 100 | $99 \%$ |
| 30 | 1 in 1000 | $99.9 \%$ |
| 40 | 1 in 10000 | $99.99 \%$ |
| 50 | 1 in 100000 | $99.999 \%$ |

## GigAssembler: Build merged sequence contigs ("rafts")



Figure 2 Two sequences with tails. The nonaligning regions on either side can be classified into 'extensions' and 'tails.' Short tails are fairly common even when two sequences should be joined into a contig because of poor quality sequence near the ends and occasional chimeric reads. Long tails, however, are generally a sign that the alignment is merely due to the sequences sharing a repeating element.

## GigAssembler: Build merged sequence contigs ("rafts")



Figure 3 Merging into a raft. A contig ('raft') of three sequences: A, $B$, and $C$ has already been constructed by GigAssembler. The program now examines an alignment between sequence $C$ and a new sequence, D, to see whether D should also be added to the raft. The parts of $D$ marked with $+s$ are compatible with the raft because of the C/D alignment. The program must also check that the parts of D marked with ?s are compatable with the raft by examining other alignments.

## GigAssembler: Build sequenced clone contigs ("barges")



Figure 4 Three overlapping draft clones: A, B, and C. Each clone has two initial sequence contigs. Note that initial sequence contigs a1, b1, and a2 overlap as do b2 and c1.

## GigAssembler: Build a "raft-ordering" graph



Figure 4 Three overlapping draft clones: A, B and C. Each colse Figure 4 Three overlapping draft clones: A, B, and C. Each clone
has two initial sequence contigs. Note that initial sequence contigs a1, b1, and a2 overlap as do b2 and c1.


Figure 6 Ordering graph after adding in rafts. The initial sequence contigs shown in Fig. 4 are merged into rafts where they overlap. This forms three rafts: albla2, b2c1, and c2. These rafts are constrained to ie between the relevant clone ends by the addition of additional ordering edges to the graph shown in Fig. 5

## GigAssembler: Build a "raft-ordering" graph

- Add information from mRNAs, ESTs, paired plasmid reads, BAC end pairs: building a "bridge"
- Different weight to different data type: (mRNA ~ highest)
- Conflicts with the graph as constructed so far are rejected.
- Build a sequence path through each raft.
- Fill the gap with N's.
- 100: between rafts
- 50,000: between bridged barges


Figure 6 Ordering graph after adding in rafts. The initial sequence contigs shown in Fig. 4 are merged into rafts where they overlap. This forms three rafts: a1b1a2, b2c1, and c2. These rafts are constrained to lie between the relevant clone ends by the addition of additional ordering edges to the graph shown in Fig. 5.

Finding the shortest path across the ordering graph using the Bellman-Ford algorithm

Find the shortest path to all nodes.
Take every edge and try to relax it ( $N-1$ times where $N$ is the count of nodes)


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## Once a reference genome is assembled, new sequencing data can 'simply' be mapped to the reference.



## Mapping reads to assembled genomes

Table 1 A selection of short-read analysis software

| Program | Website | Open <br> source? | Handles ABI color <br> space? | Maximum read <br> length |
| :--- | :--- | :---: | :---: | :---: |
| Bowtie | http://bowtie.cbcb.umd.edu | Yes | No | None |
| BWA | http://maq.sourceforge.net/bwa-man.shtml | Yes | Yes | None |
| Maq | http://maq.sourceforge.net | Yes | Yes | 127 |
| Mosaik | http://bioinformatics.bc.edu/marthlab/Mosaik | No | Yes | None |
| Novoalign | http://www.novocraft.com | No | No | None |
| SOAP2 | http://soap.genomics.org.cn | No | No | 60 |
| ZOOM | http://www.bioinfor.com | No | Yes | 240 |




## Burroughs-Wheeler transform indexing

BWT is often used for file compression (like bzip2), here used to make a fast 'lookup' index in a genome

BWT = 'reversible block-sorting'

Input SIX.MIXED.PIXIES.SIFT.SIXTY.PIXIE.DUST.BOXES
Forward BWT $\quad$ This sequence is
Output TEXYDST.E.IXIXIXXSSMPPS.B..E.S.EUSFXDIIOIIIT
Reverse BWT
Recovered SIX.MIXED.PIXIES.SIFT.SIXTY.PIXIE.DUST.BOXES input



## Burroughs-Wheeler transform indexing

## Sorting All Rows in Alphabetical Order

ANANA| ${ }^{\wedge} B$
ANA|^BAN
A|^BANAN
BANANA|^
NANA|^BA
NA|^BANA
${ }^{\wedge}$ BANANA|
|^BANANA

## Burroughs-Wheeler transform indexing

| Taking Last Column |
| :---: |
| ANANA ${ }^{\wedge}$ B |
| ANA\|^BAN |
| A ${ }^{\wedge}$ BANAN |
| BANANA\|^ |
| NANA ${ }^{\wedge}$ BA |
| NA\|^BANA |
| ^BANANA |
| $l^{\wedge}$ BANANA |

## Burroughs-Wheeler transform indexing

| Output <br> Last Column |
| :---: |
|  |
|  |
| BNN^AA $\mid A$ |
|  |

## Burroughs-Wheeler transform indexing

| Transformation |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Input | All <br> Rotations | Sorting All Rows in Alphabetical Order | Taking Last Column | Output Last Column |
| ^BANANA | ^BANANA\| <br> $\left.\right\|^{\wedge}$ BANANA <br> A\| ${ }^{\wedge}$ BANAN <br> NA ${ }^{\wedge}$ BANA <br> ANA\|^BAN <br> NANA\| ${ }^{\wedge} \mathrm{BA}$ <br> ANANA\|^B <br> BANANA\|^ | ANANA $\left.\right\|^{\wedge}$ B <br> ANA\|^BAN <br> A ${ }^{\wedge}$ BANAN <br> BANANA\|^ <br> NANA\| ^BA <br> NA\| ${ }^{\text {BANA }}$ <br> ${ }^{\wedge}$ BANANA\| <br> $I^{\wedge}$ BANANA | ANANA ${ }^{\wedge}$ •B <br> ANA\|^BAN <br> A\|^BANAN <br> BANANA\|^ <br> NANA\|^BA <br> NA\| ${ }^{\text {BANA }}$ <br> ^BANANA। <br> $\left.\right\|^{\wedge}$ BANANA | BNN^AA $\mid$ A |

## BWT is remarkable because it is reversible.

Any ideas as how you might reverse it?



## Burroughs-Wheeler transform indexing

| Add 3 | Sort 3 | Add 4 | Sort 4 |
| :---: | :---: | :---: | :---: |
| BAN | ANA | BANA | ANAN |
| NAN | ANA | NANA | ANA 1 |
| NA 1 | A ${ }^{\wedge}$ | NA ${ }^{\wedge}$ | $A \mid \wedge B$ |
| $\wedge$ BA | BAN | $\wedge$ BAN | BANA |
| ANA | NAN | ANAN | NANA |
| ANA | NA ! | ANA 1 | NA $\left.\right\|^{\wedge}$ |
| $\\|^{\wedge} \mathrm{B}$ | $\wedge$ BA | $\\|^{\wedge} \mathrm{BA}$ | ^BAN |
| $\left.\mathrm{A}\right\|^{\wedge}$ | $\\|^{\wedge} \mathrm{B}$ | $A \mid \wedge B$ | $\\|^{\wedge} \mathrm{BA}$ |
| Add the columns... | Sort those... | Add the columns... | Sort those... |


| Burroughs-Wheeler transform indexing |  |  |  |
| :---: | :---: | :---: | :---: |
| Add 5 | Sort 5 | Add 6 | Sort 6 |
| BANAN | ANANA | BANANA | ANANA |
| NANA | ANA\|^ | NANA\|^ | ANA ${ }^{\wedge} \mathrm{B}$ |
| NA ${ }^{\wedge} \mathrm{B}$ | A ${ }^{\wedge} \mathrm{BA}$ | NA ${ }^{\wedge} \mathrm{BA}$ | Al^BAN |
| ${ }^{\wedge}$ BANA | BANAN | ^BANAN | BANANA |
| ANANA | NANA I | ANANA | NANA ${ }^{\wedge}$ |
| ANA\|^ | NA ${ }^{\wedge}$ ^B | ANA $\wedge^{\wedge} \mathrm{B}$ | NA $\wedge^{\wedge} \mathrm{BA}$ |
| $\\|^{\wedge} \mathrm{BAN}$ | ^BANA | $\\|^{\wedge}$ BANA | ^BANAN |
| $\left.\mathrm{A}\right\|^{\wedge} \mathrm{BA}$ | $\\|^{\wedge}$ BAN | A ${ }^{\wedge} \mathrm{BAN}$ | $\\|^{\wedge}$ BANA |
| Add the columns... | Sort those... | Add the columns... | Sort those... |

## Burroughs-Wheeler transform indexing

| Add 7 | Sort 7 | Add 8 |  |
| :---: | :---: | :---: | :---: |
| BANANA | ANANA\|^ | BANANA\|^ |  |
| NANA ${ }^{\wedge}$ ^B | ANA ${ }^{\wedge} \mathrm{BA}$ | NANA\|^BA | The row with |
| NA ${ }^{\wedge}$ ^BAN | A ${ }^{\wedge}$ BANA | NA ${ }^{\wedge}$ BANA | the "end of file" |
| $\wedge$ BANANA | BANANA | ABANANAD | character at the |
| ANANA\|^ | NANA ${ }^{\wedge} \mathrm{B}$ | ANANA ${ }^{\wedge}$ B | end is the |
| ANA ${ }^{\wedge}$ BA | NA\|^BAN | ANA\|^BAN | original text |
| \|^BANAN | ^BANANA | $\\|^{\wedge}$ BANANA |  |
| A ${ }^{\wedge}$ BANA | $\\|^{\wedge}$ BANAN | A ${ }^{\wedge}$ BANAN |  |
| Add the | Sort those... | Add the |  |


| Burroughs-Wheeler transform indexing |
| :---: |
| Output |
| ${ }^{\wedge}$ BANANA । |
| The row with the "end of file" <br> character at the end is the <br> original text |
| nttp://en.wikipedia.org/wiki/Burrows-Wheeler transform |


|  | Burroughs Wheeler indexing |
| :---: | :---: |

