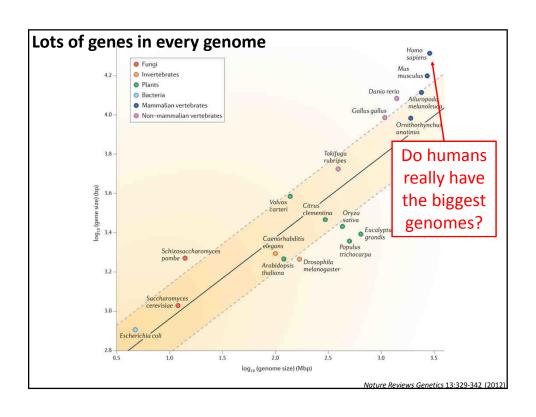
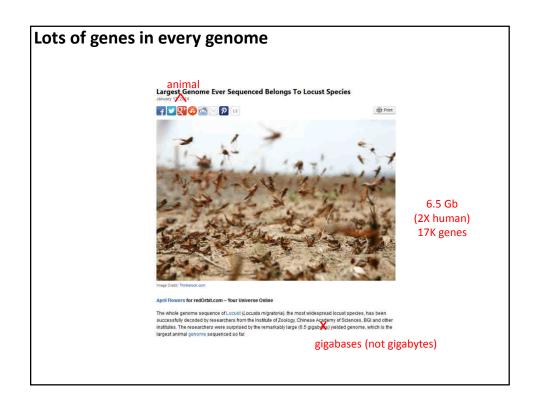
Gene Finding

BCH339N Systems Biology / Bioinformatics Edward Marcotte, Univ of Texas at Austin

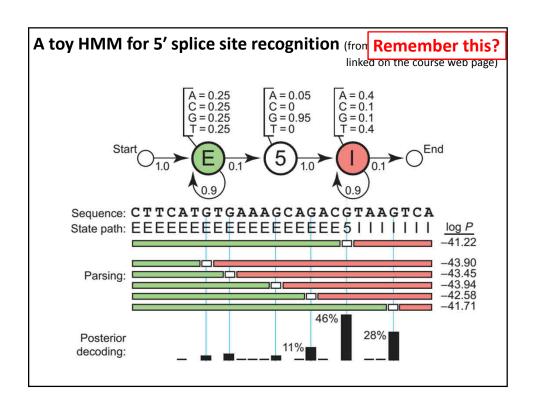






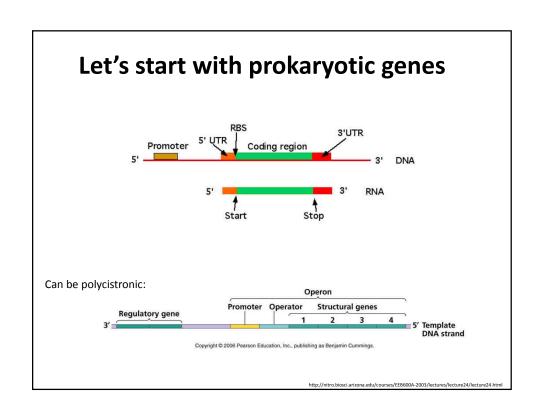
Where are the genes? How can we find them?

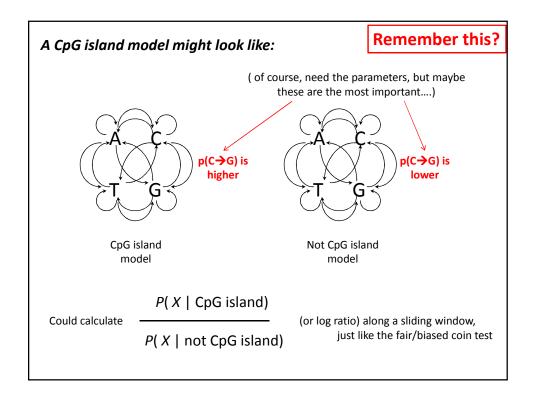
GATCACTTGATAAATGGGCTGAAGTAACTCGCCCAGATGAGGAGTGTGCTGCCTCCAGAAT CCAAACAGGCCCACTAGGCCCGAGACACCTTGTCTCAGATGAAACTTTGGACTCGGAATT TTGAGTTAATGCCGGAATGAGTTCAGACTTTGGGGGACTGTTGGGAAGGCATGATTGGTT TCAAAATGTGAGAAGGACATGAGATTTGGGAGGGGCTGGGGGCAGAATGATATAGTTTG GCTCTGCGTCCCCACCCAATCTCATGTCAAATTGTAATCCTCATGTGTCAGGGGAGAGGCCT GGTGGGATGTGATTGGATCATGGGAGTGGATTTCCCTCTTGCAGTTCTCGTGATAGTGAGT TCCCCTGCTCCACCATGGTGAGACGTGCTTGCGTCCCCTTTGCCTTCTGCCATGATTGTAAG CTTCCTCAGGCGTCCTAGCCACGCTTCCTGTACAGCCTGAGGAACTGGGAGTCAATGAAA TACAAGTAGAGACTGAGATCAATAGCATTTGCACTGGGCCTGGAACACACTGTTAAGAAC GTAAGAGCTATTGCTGTCATTAGTAATATTCTGTATTATTGGCAACATCACACAATACACTGC TGTGGGAGGGTCTGAGATACTTCTTTGCAGACTCCAATATTTGTCAAAACATAAAATCAGG AGCCTCATGAATAGTGTTTAAATTTTTACATAATAATACATTGCACCATTTGGTATATGAGTCT TTTTGAAATGGTATATGCAGGACGGTTTCCTAATATACAGAATCAGGTACACCTCCTCTTCCA TACTGCTGTTGTTACCCACAGTGCACCTCAGACTCACGTTTCTCCAGCAATGAGCTCCTGTT CCCTGCACTTAGAGAAGTCAGCCCGGGGACCAGACGGTTCTCTCCTCTTGCCTGCTCCAG CCTTGGCCTTCAGCAGTCTGGATGCCTATGACACAGAGGGCATCCTCCCCAAGCCCTGGTC CTTCTGTGAGTGGTGAGTTGCTGTTAATCCAAAAGGACAGGTGAAAACATGAAAGCC...

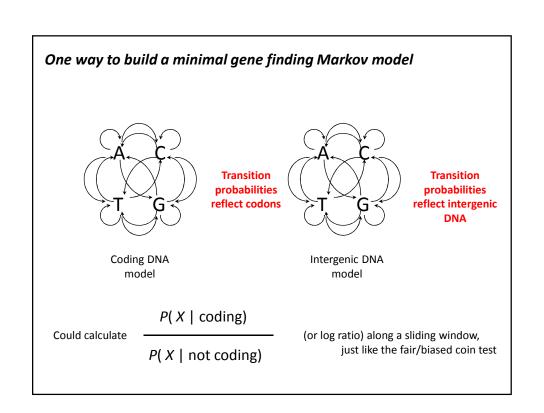


Let's start with prokaryotic genes

What elements should we build into an HMM to find bacterial genes?





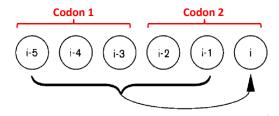


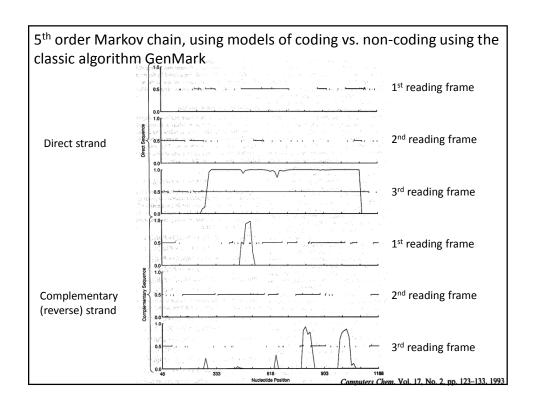
Really, we'll want to detect codons.

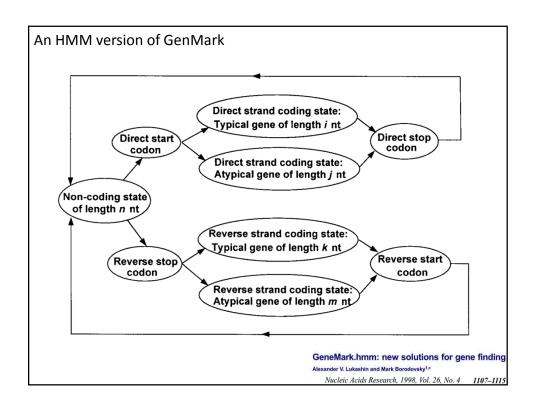
The usual trick is to use a higher-order Markov process.

A standard Markov process only considers the current position in calculating transition probabilities.

An n^{th} -order Markov process takes into account the past n nucleotides, e.g. as for a 5th order:





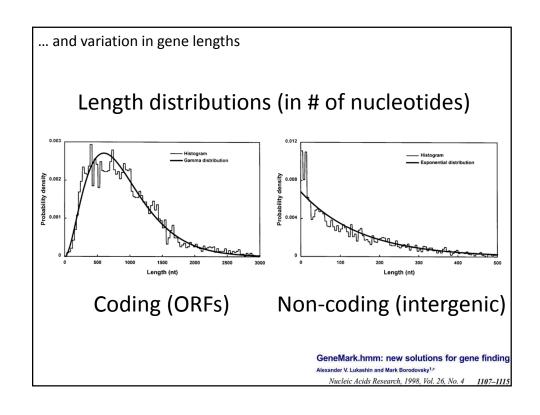


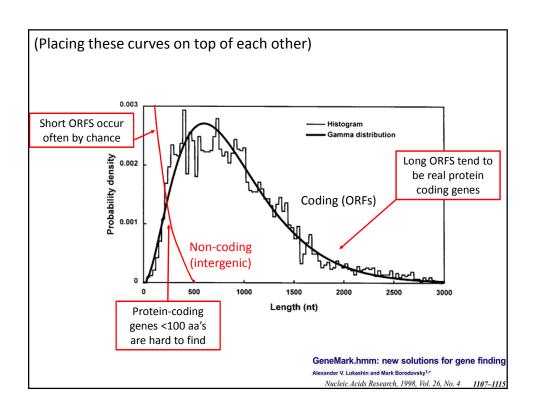
For example, accounting for variation in start codons...

The probabilities of the start codons were defined in agreement with the *E.coli* genome statistics: P(ATG) = 0.905, P(GTG) = 0.090, P(TTG) = 0.005. The probability of transition from a non-coding state to a Typical (Atypical) coding state was set to 0.85 (0.15).

GeneMark.hmm: new solutions for gene finding
Alexander V. Lukashin and Mark Borodovsky^{1,4}

Nucleic Acids Research, 1998, Vol. 26, No. 4 1107–1115





Model for a ribosome binding site (based on ~300 known RBS's)

Nucleotide	Position					
	1	2	3	4	5	
T	0.161	0.050	0.012	0.071	0.115	
C	0.077	0.037	0.012	0.025	0.046	
A	0.681	0.105	0.015	0.861	0.164	
G	0.077	0.808	0.960	0.043	0.659	

GeneMark.hmm: new solutions for gene finding
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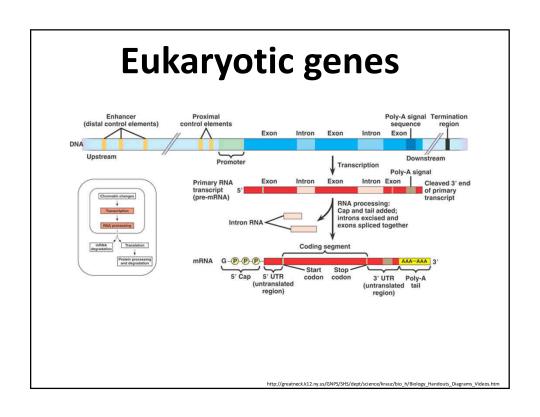
How well does it do on well-characterized genomes?

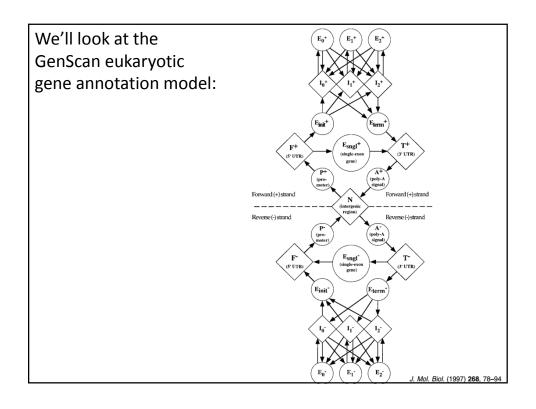
Genome	Genes annotated	Genes predicted	Exact prediction (%)	Missing genes (%)	Wrong genes (%)
A.fulgidus	2407	2530	73.1	10.8 (2.0)	15.1
B.subtilis	4101	4384	77.5	3.6 (2.8)	9.8
E.coli	4288	4440	75.4	5.0 (2.7)	8.2
H.influenzae	1718	1840	86.7	3.8 (3.2)	10.2
H.pylori	1566	1612	79.7	6.0 (4.4)	8.7
M.genitalium	467	509	78.4	9.9 (1.7)	17.3
M.jannaschii	1680	1841	72.7	4.6 (0.8)	12.9
M.pneumoniae	678	734	70.1	7.8 (4.1)	13.6
M.thermoauthotrophicum	1869	1944	70.9	5.0 (3.5)	8.6
Synechocystis	3169	3360	89.6	4.0 (1.5)	9.4
Averaged	21 943	23 194	78.1	5.4 (2.7)	10.4

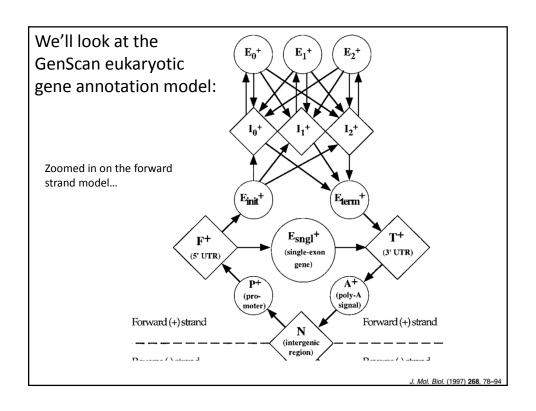
But this was a long time ago!

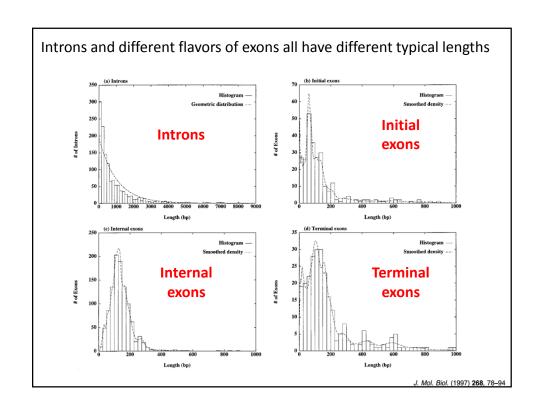
Eukaryotic genes

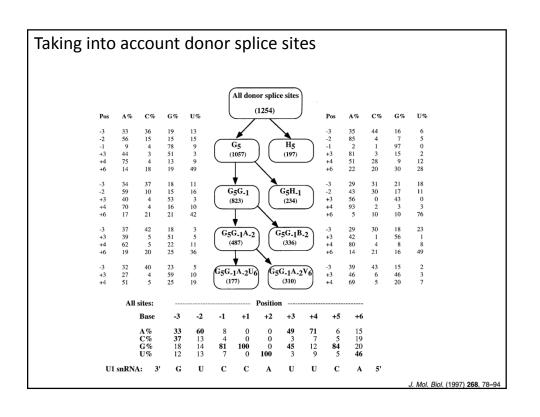
What elements should we build into an HMM to find eukaryotic genes?

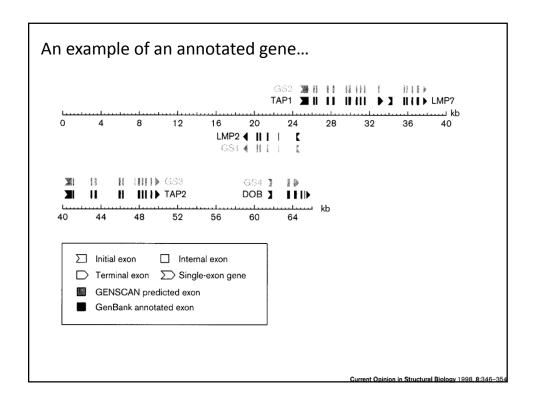


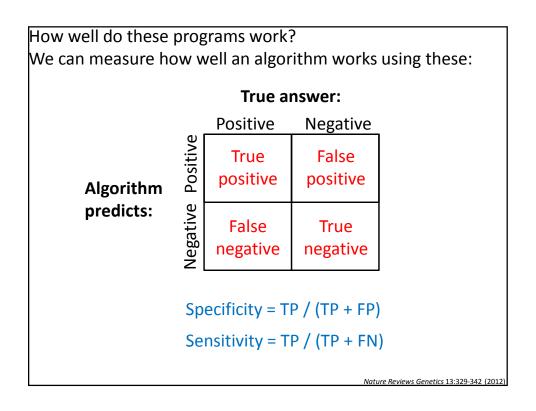


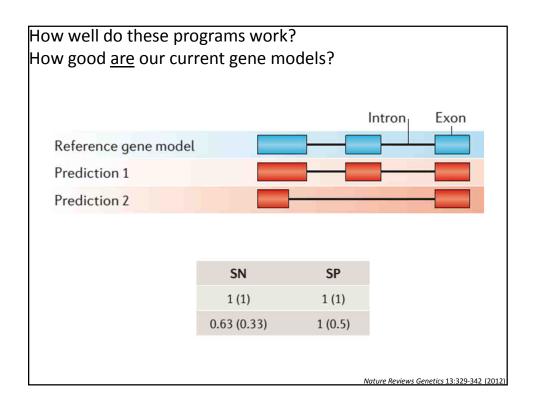




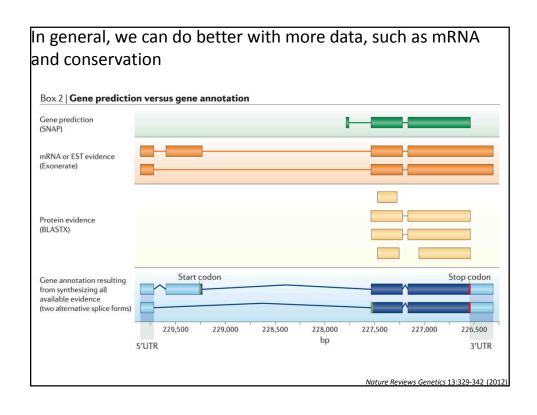


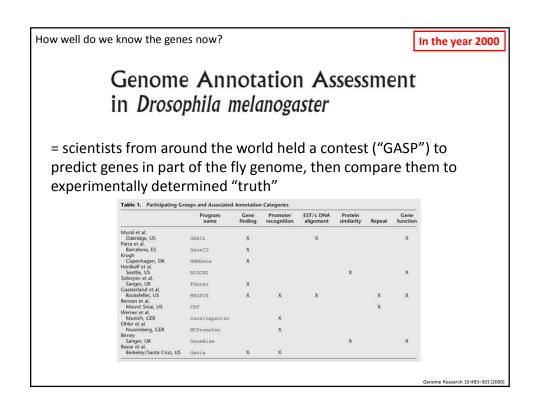






		Accuracy per base		Accuracy per exon	
	Sequences				
Program		Sn	Sp	Sn	Sp
GENSCAN	570 (8)	0.93	0.93	0.78	0.81
FGENEH	569 (22)	0.77	0.88	0.61	0.64
GeneID	570 (2)	0.63	0.81	0.44	0.46
Genie	570 (0)	0.76	0.77	0.55	0.48
GenLang	570 (30)	0.72	0.79	0.51	0.52
GeneParser2	562 (0)	0.66	0.79	0.35	0.40
GRAIL2	570 (23)	0.72	0.87	0.36	0.43
SORFIND	561 (0)	0.71	0.85	0.42	0.47
Xpound	570 (28)	0.61	0.87	0.15	0.18
GeneID+	478 (1)	0.91	0.91	0.73	0.70
GeneParser3	478 (1)	0.86	0.91	0.56	0.58





In the year 2000

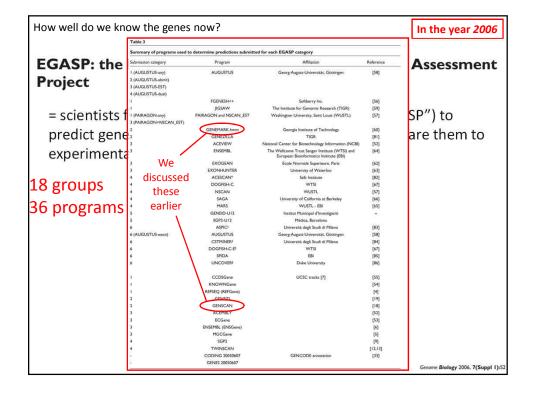
"Over <u>95%</u> of the coding nucleotides ... were correctly identified by the majority of the gene finders."

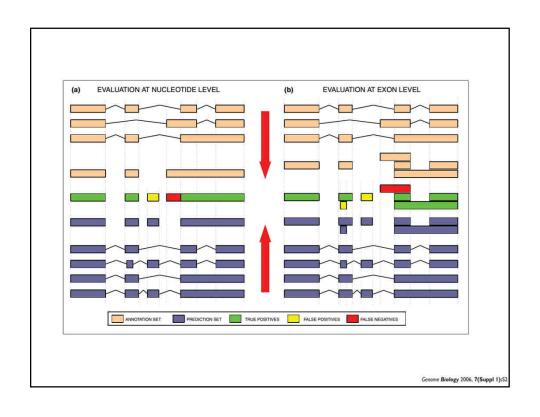
"...the correct intron/exon structures were predicted for \geq 40% of the genes."

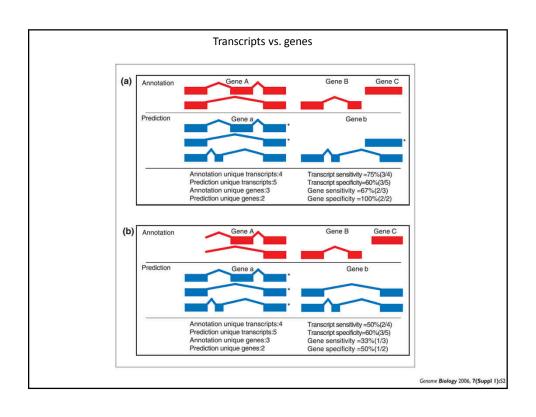
Most promoters were missed; many were wrong.

"Integrating gene finding and cDNA/EST alignments with promoter predictions decreases the number of false-positive classifications but <u>discovers less than one-third of the</u> promoters in the region."

Genome Research 10:483-501 (200





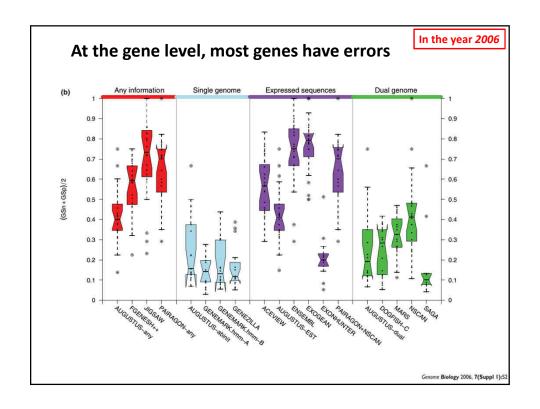


In the year 2006

So how did they do?

- "The best methods had <u>at least one gene transcript</u> correctly predicted for close to **70%** of the annotated genes."
- "...taking into account alternative splicing, ... only approximately **40**% to **50**% accuracy.
- At the coding <u>nucleotide</u> level, the best programs reached an accuracy of **90%** in both sensitivity and specificity."

Genome Biology 2006, 7(Suppl 1):S



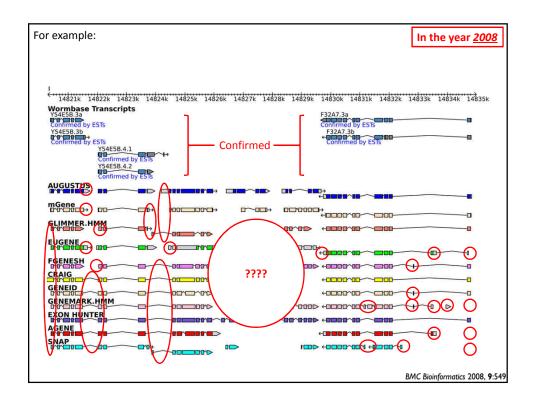
In the year <u>2008</u>

nGASP - the nematode genome annotation assessment project

= scientists from around the world held a contest ("NGASP") to predict genes in part of the <u>worm</u> genome, then compare them to experimentally determined "truth"

- · 17 groups from around the world competed
- "Median gene level sensitivity ... was 78%"
- "their specificity was 42%", comparable to human

BMC Bioinformatics 2008, 9:549



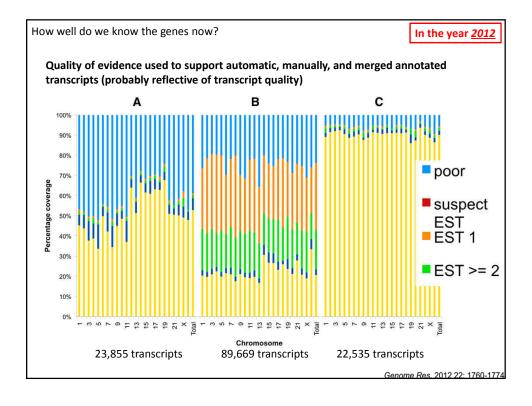
In the year <u>2012</u>

GENCODE: The reference human genome annotation for The ENCODE Project

= a large consortium of scientists trying to annotate the <u>human</u> genome using a combination of experiment and prediction.

Best estimate of the current state of human genes.

Genome Res. 2012 22: 1760-1774



In the year <u>2015</u>

The bottom line:

- Gene prediction and annotation are hard
- · Annotations for all organisms are still buggy
- Few genes are 100% correct; expect multiple errors per gene
- Most organisms' gene annotations are probably much worse than for humans

