Assembling Genomes

BCH364C/394P Systems Biology / Bioinformatics
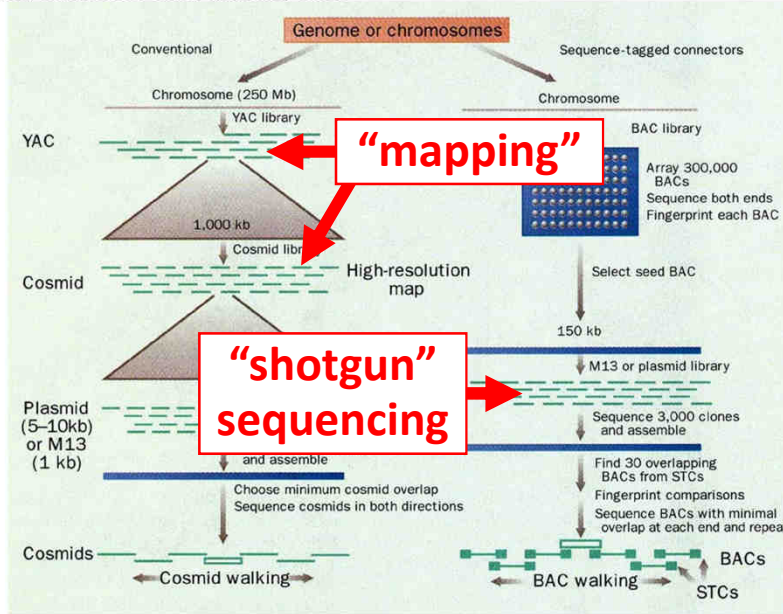
Edward Marcotte, Univ of Texas at Austin

A new strategy for genome sequencing

J. Craig Venter, Hamilton O. Smith and Leroy Hood

"mapping"

"shotgun" sequencing

(Translating the cloning jargon)

CLONE LIBRARIES USED FOR GENOME MAPPING AND SEQUENCING

Vector	Human-DNA insert size range	Number of clones required to cover the human genome
Yeast artificial chromosome (YAC)	100–2,000 kb	3,000 (1,000 kb)
Bacterial artificial chromosome (BAC)	80–350 kb	20,000 (150 kb)
Cosmid	30–45 kb	75,000 (40 kb)
Plasmid	3–10 kb	600,000 (5 kb)
M13 phage	1 kb	3,000,000 (1 kb)

Thinking about the basic shotgun concept

- Start with a very large set of random sequencing reads

- How might we match up the overlapping sequences?

- How can we assemble the overlapping reads together in order to derive the genome?

Thinking about the basic shotgun concept

- At a high level, the first genomes were sequenced by comparing pairs of reads to find overlapping reads

- Then, building a graph (*i.e.*, a network) to represent those relationships

- The genome sequence is a "walk" across that graph

The "Overlap-Layout-Consensus" method

Overlap: Compare all pairs of reads
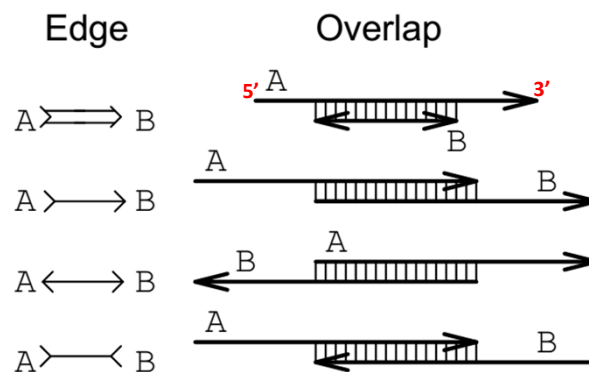(allow some low level of mismatches)

Layout: Construct a graph describing the overlaps

sequence
overlap
read
read
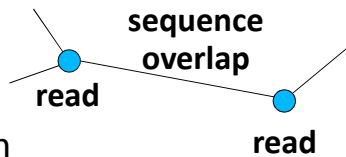
Simplify the graph
Find the simplest path through the graph

Consensus: Reconcile errors among reads along that
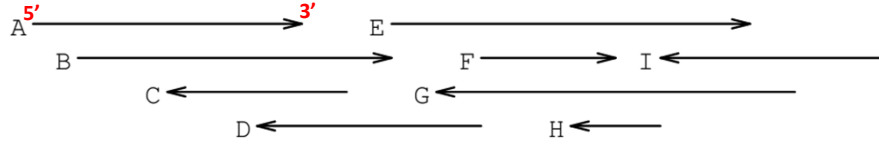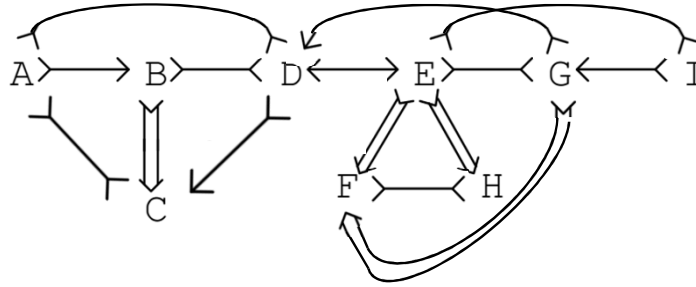path to find the consensus sequence

---

Building an overlap graph

Edge   Overlap

A ⟹ B

A ⟩—→ B

A ⟵—→ B

A ⟩—⟨ B

Building an overlap graph

Reads

Overlap graph

Simplifying an overlap graph
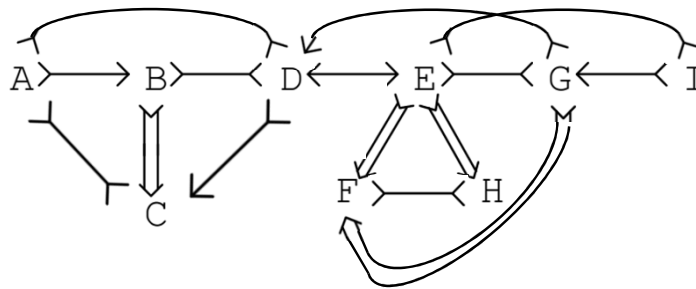
1. Remove all contained nodes & edges going to them

Simplifying an overlap graph

A ⟩—→ B ⟩—⟨ D ⟷ E ⟩—⟨ G ⟵—⟨ I
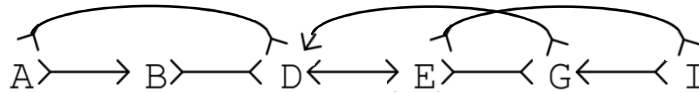
2. Transitive edge removal:
   Given A – B – D   and   A – D , remove A – D

Simplifying an overlap graph

A ⟩—→ B ⟩—⟨ D ⟷ E ⟩—⟨ G ⟵—⟨ I
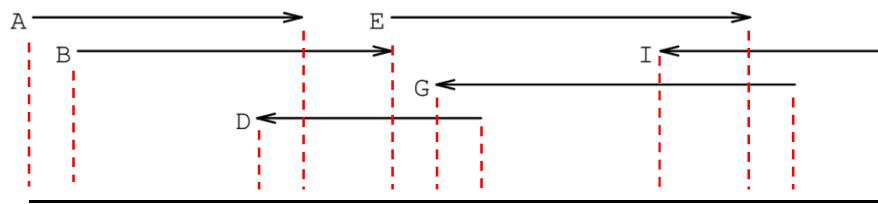
3. If un-branched, calculate consensus sequence
   If branched, assemble un-branched bits and then decide
      how they fit together

Simplifying an overlap graph

A >——> B >——< D <——> E >——< G <——< I

A ————————→
  B ————————→
      D <————
          G <————
  E ————————→
      I <————

"contig" (assembled contiguous sequence)

This basic strategy was used for most of the early genomes.
Also useful: "mate pairs"

2 reads separated by a known distance

Read #1

DNA fragment of known size

Read #2
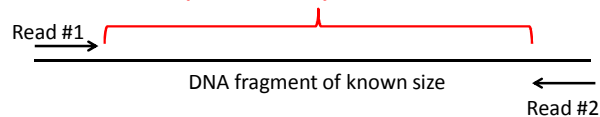
Contigs can be ordered using these paired reads

Contig #1   Contig #2

to produce "scaffolds"

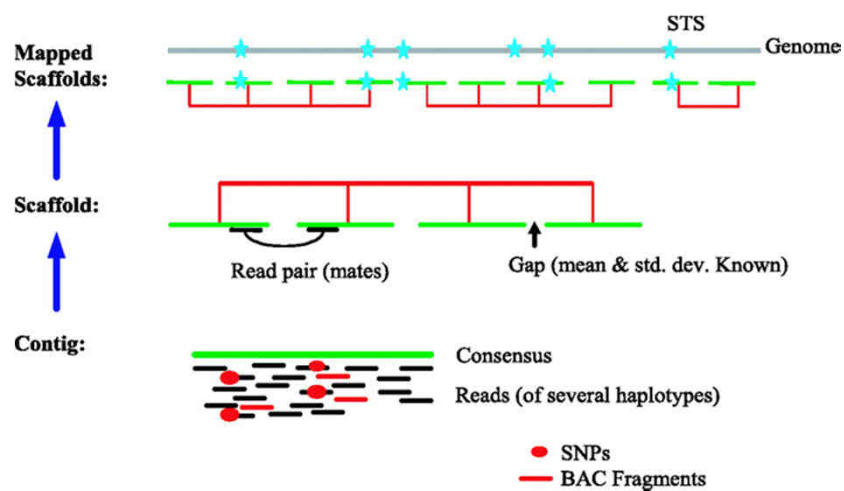# GigAssembler (used to assemble the public human genome project sequence)

Jim Kent   David Haussler

Whole genome Assembly: big picture

http://www.nature.com/scitable/content/anatomy-of-whole-genome-assembly-20429

GigAssembler – Preprocessing

1. Decontaminating & Repeat Masking.

2. Aligning of mRNAs, ESTs, BAC ends & paired reads against initial sequence contigs.

   - psLayout → BLAT

3. Creating an input directory (folder) structure.

RepBase + RepeatMasker

GigAssembler: Build merged sequence contigs ("rafts")

Figure 1 Two sequences overlapping end to end. The sequences are represented as dashes. The aligning regions are joined by vertical bars. End-to-end overlap is an extremely strong indication that two sequences should be joined into a contig.

Sequencing quality (Phred Score)

Sequencing quality (Phred Score)

$$Q = -10 \ \log_{10} P$$ ← *Base-calling Error Probability*

or

$$P = 10^{\frac{-Q}{10}}$$

Phred quality scores are logarithmically linked to error probabilities

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90 %
20	1 in 100	99 %
30	1 in 1000	99.9 %
40	1 in 10000	99.99 %
50	1 in 100000	99.999 %

http://en.wikipedia.org/wiki/Phred_quality_score

GigAssembler: Build merged sequence contigs ("rafts")

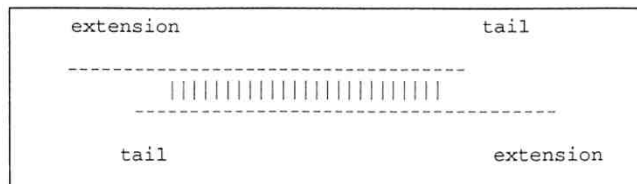**Figure 2** Two sequences with tails. The nonaligning regions on either side can be classified into 'extensions' and 'tails.' Short tails are fairly common even when two sequences should be joined into a contig because of poor quality sequence near the ends and occasional chimeric reads. Long tails, however, are generally a sign that the alignment is merely due to the sequences sharing a repeating element.

GigAssembler: Build merged sequence contigs ("rafts")

```
A -----------------
     B -------------------
   C-------------
              D ++++++?????????--------------
```
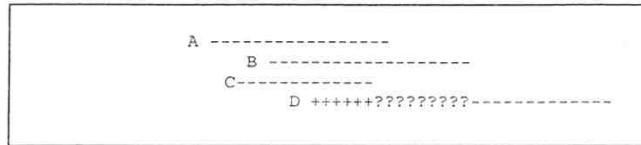
Figure 3 Merging into a raft. A contig ('raft') of three sequences: A, B, and C has already been constructed by GigAssembler. The program now examines an alignment between sequence C and a new sequence, D, to see whether D should also be added to the raft. The parts of D marked with +s are compatible with the raft because of the C/D alignment. The program must also check that the parts of D marked with ?s are compatable with the raft by examining other alignments.

GigAssembler: Build sequenced clone contigs ("barges")
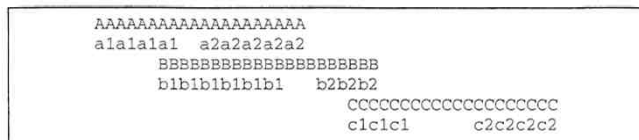
```
AAAAAAAAAAAAAAAAAAAA
a1a1a1a1   a2a2a2a2a2
      BBBBBBBBBBBBBBBBBBBB
    b1b1b1b1b1b1    b2b2b2
                      CCCCCCCCCCCCCCCCCCCCCC
                    c1c1c1        c2c2c2c2
```

Figure 4 Three overlapping draft clones: A, B, and C. Each clone has two initial sequence contigs. Note that initial sequence contigs a1, b1, and a2 overlap as do b2 and c1.

GigAssembler:
Build a "raft-ordering" graph

Figure 4 Three overlapping draft clones: A, B, and C. Each clone has two initial sequence contigs. Note that initial sequence contigs a1, b1, and a2 overlap as do b2 and c1.

Figure 5 Ordering graph of clone starts and ends. This represents the same clones as in Fig. 4. (As) The start of clone A; (Ae) the end of clone A. Similarly Bs, Be, Cs, and Ce represent the starts and ends of clones B and C.
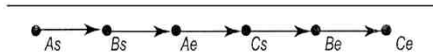
Figure 6 Ordering graph after adding in rafts. The initial sequence contigs shown in Fig. 4 are merged into rafts where they overlap. This forms three rafts: a1b1a2, b2c1, and c2. These rafts are constrained to lie between the relevant clone ends by the addition of additional ordering edges to the graph shown in Fig. 5.

GigAssembler:
Build a "raft-ordering" graph

- Add information from mRNAs, ESTs, paired plasmid reads, BAC end pairs: building a "bridge"
  - Different weight to different data type: (mRNA ~ highest)
  - Conflicts with the graph as constructed so far are rejected.
- Build a sequence path through each raft.
- Fill the gap with N's.
  - 100: between rafts
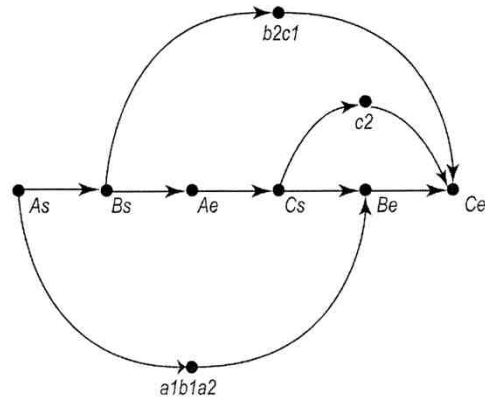 - 50,000: between bridged barges
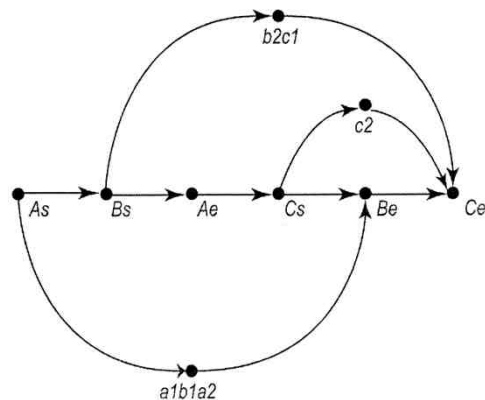
Figure 6 Ordering graph after adding in rafts. The initial sequence contigs shown in Fig. 4 are merged into rafts where they overlap. This forms three rafts: a1b1a2, b2c1, and c2. These rafts are constrained to lie between the relevant clone ends by the addition of additional ordering edges to the graph shown in Fig. 5.

Finding the shortest path across the
ordering graph using the
Bellman-Ford algorithm

Find the shortest path to all nodes.

Take every edge and try to relax it (N – 1 times where N is the count of nodes)

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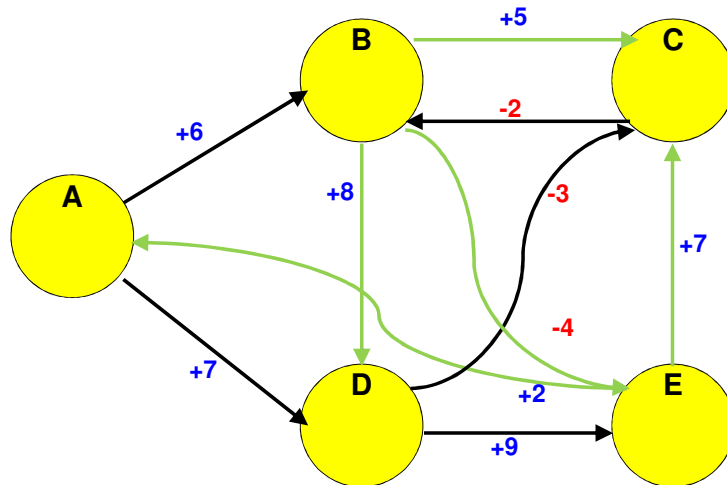

Find the shortest path to all nodes.

Take every edge and try to relax it (N – 1 times where N is the count of nodes)

17

Answer: A-D-C-B-E
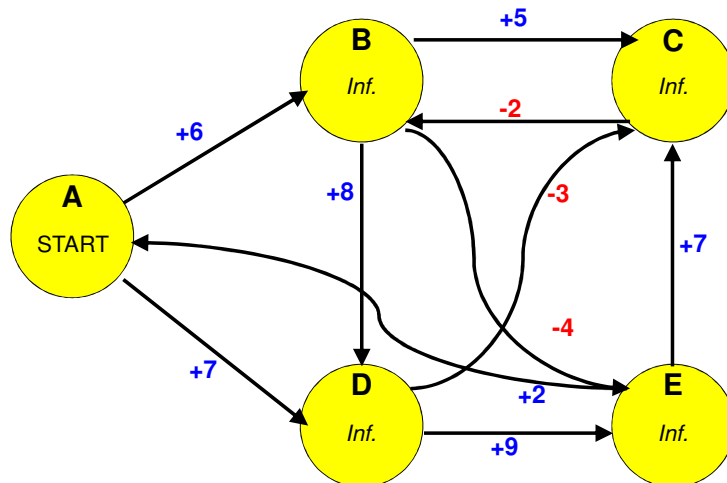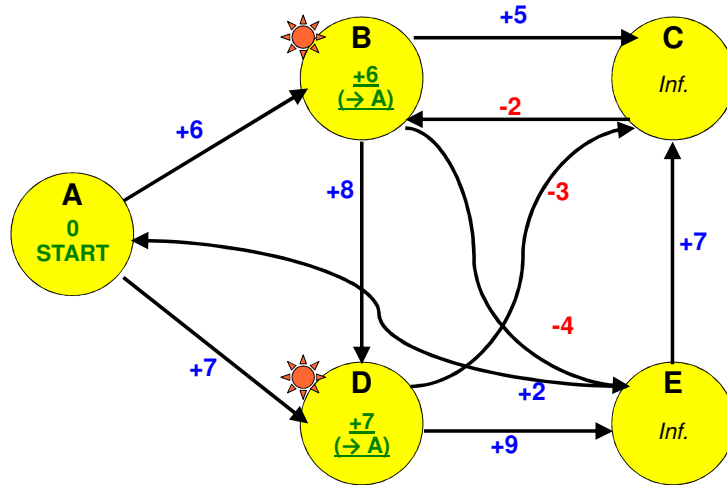

Modern assemblers now work a bit differently, using so-called DeBruijn graphs:

Here's what we saw before:
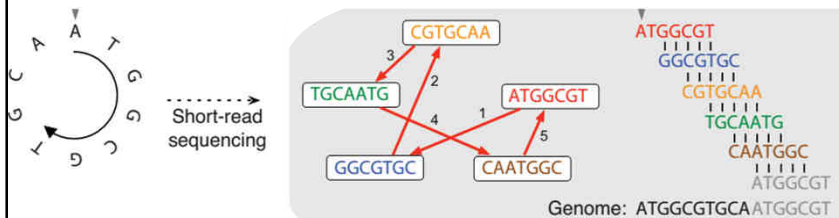
In Overlap-Layout-Consensus:
Nodes are reads
Edges are overlaps

Modern assemblers now work a bit differently, using so-called DeBruijn graphs:

In a DeBruijn graph:

Vertices are (k–1)-mers
Edges are k-mers

Genome: ATGGCGTGCAATG

Eulerian cycle
Visit each edge once

Nature Biotech 29(11):987-991 (2011)
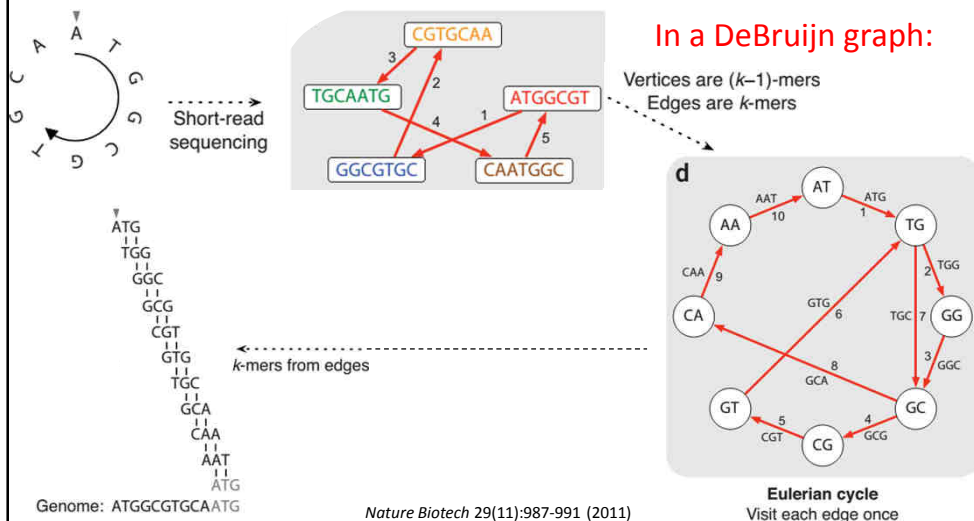
Why Eulerian?

From Leonhard Euler's solution in 1735 to the
'Bridges of Königsberg' problem:

Königsberg (now Kaliningrad, Russia) had 7 bridges connecting 4 parts of the city. **Could you visit each part of the city, walking across each bridge only once, & finish back where you started?**

(Visiting every edge once = an *Eulerian* path)
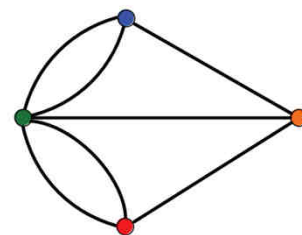
Euler conceptualized it as a graph:
Nodes = parts of city
Edges = bridges

Nature Biotech 29(11):987-991 (2011)

DeBruijn graph assemblers tend to have nice properties, e.g. correcting sequencing errors & handling repeats better

Sequencing errors appear as 'bulges'

Removing the 'bulges' corrects the errors (e.g. leaves the red path)
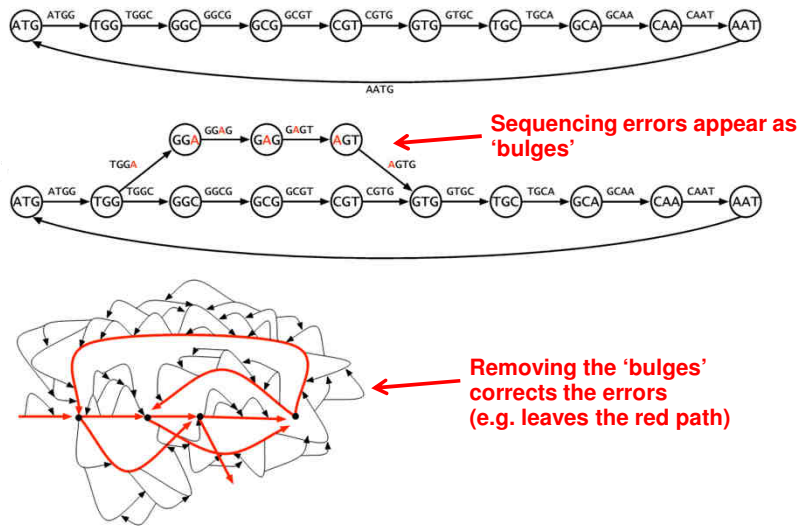
Once a reference genome is assembled, new sequencing data can 'simply' be mapped to the reference.

reads

Reference genome
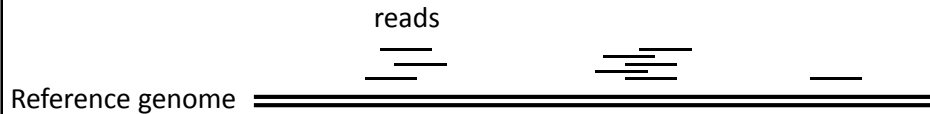
Mapping reads to assembled genomes

Table 1 A selection of short-read analysis software				
Program	Website	Open source?	Handles ABI color space?	Maximum read length
Bowtie	http://bowtie.cbcb.umd.edu	Yes	No	None
BWA	http://maq.sourceforge.net/bwa-man.shtml	Yes	Yes	None
Maq	http://maq.sourceforge.net	Yes	Yes	127
Mosaik	http://bioinformatics.bc.edu/marthlab/Mosaik	No	Yes	None
Novoalign	http://www.novocraft.com	No	No	None
SOAP2	http://soap.genomics.org.cn	No	No	60
ZOOM	http://www.bioinfor.com	No	Yes	240

The list is a little longer now! e.g. see   https://en.wikipedia.org/wiki/List_of_sequence_alignment_software#Short-Read_Sequence_Alignment

Trapnell C, Salzberg SL, *Nat. Biotech.*, 2009

Mapping strategies

Trapnell C, Salzberg SL, *Nat. Biotech.*, 2009

Burroughs Wheeler indexing

Trapnell C, Salzberg SL, *Nat. Biotech.*, 2009
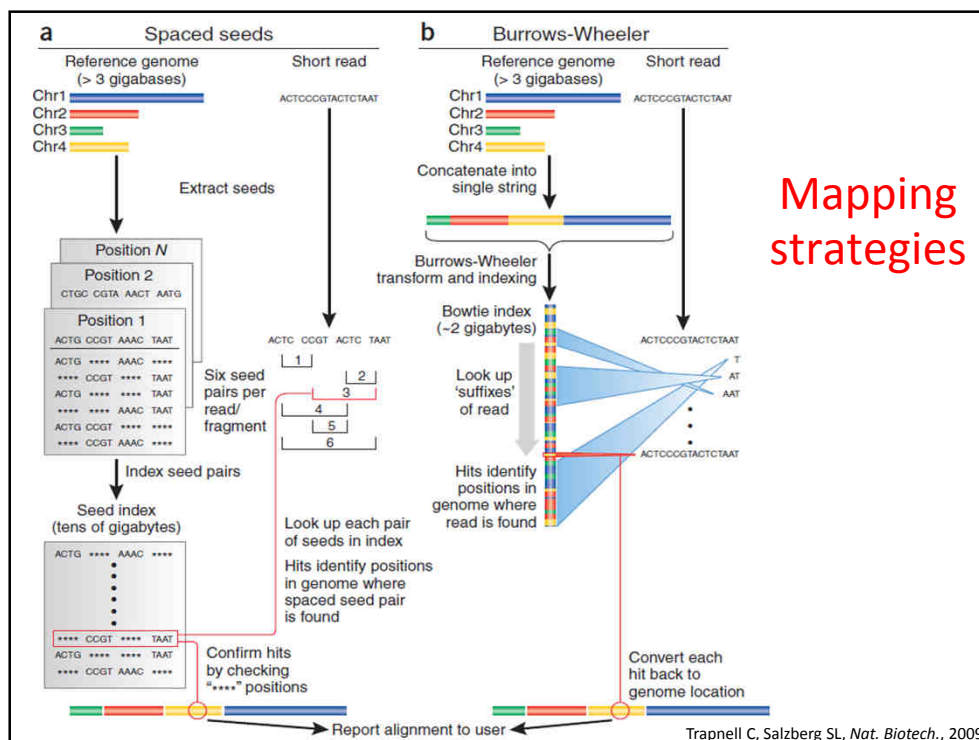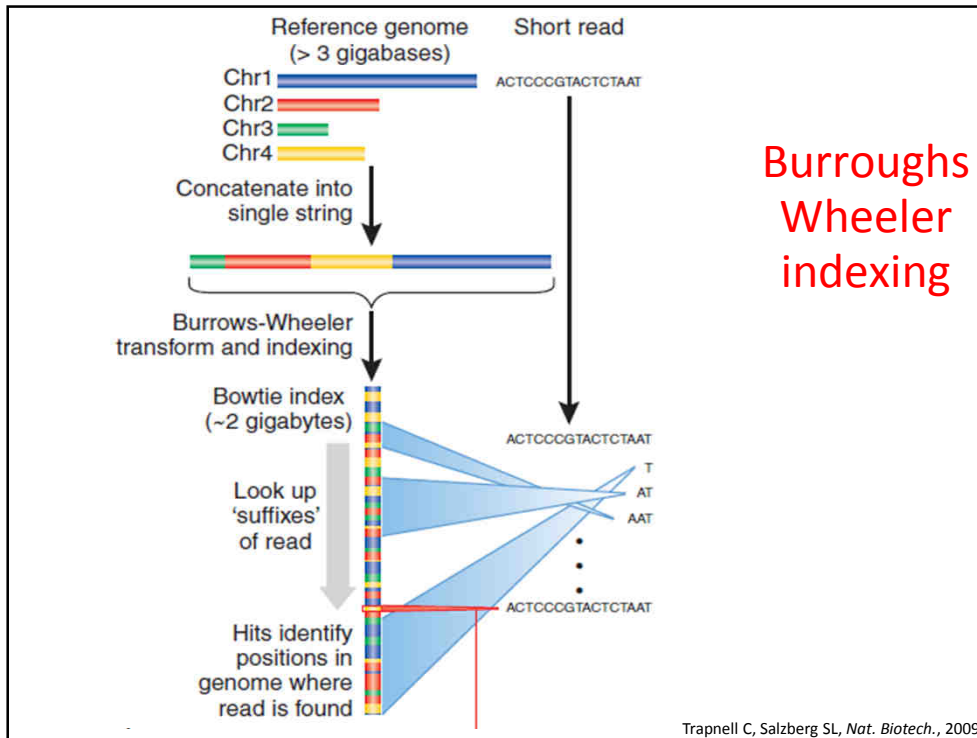
Burroughs-Wheeler transform indexing

BWT is often used for file compression (like bzip2),
here used to make a fast 'lookup' index in a genome

BWT = 'reversible block-sorting'
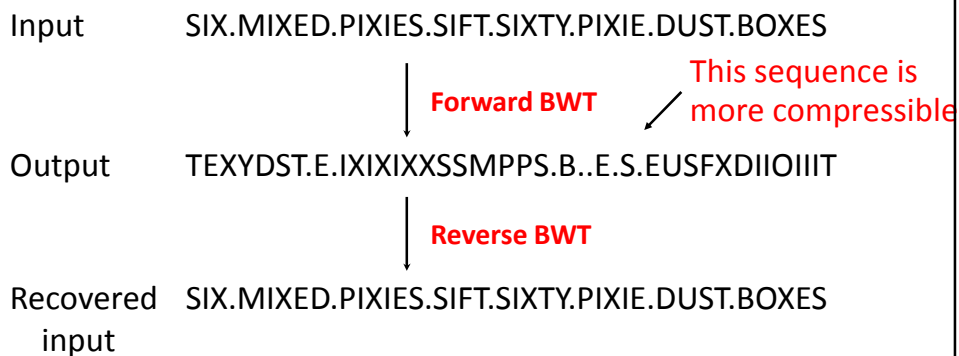
Input	SIX.MIXED.PIXIES.SIFT.SIXTY.PIXIE.DUST.BOXES	
	Forward BWT	This sequence is more compressible
Output	TEXYDST.E.IXIXIXXSSMPPS.B..E.S.EUSFXDIIOIIIT	
	Reverse BWT	
Recovered input	SIX.MIXED.PIXIES.SIFT.SIXTY.PIXIE.DUST.BOXES	

http://en.wikipedia.org/wiki/Burrows-Wheeler_transform

Burroughs-Wheeler transform indexing

Input
^BANANA|

Burroughs-Wheeler transform indexing

All Rotations
^BANANA|
|^BANANA
A|^BANAN
NA|^BANA
ANA|^BAN
NANA|^BA
ANANA|^B
BANANA|^
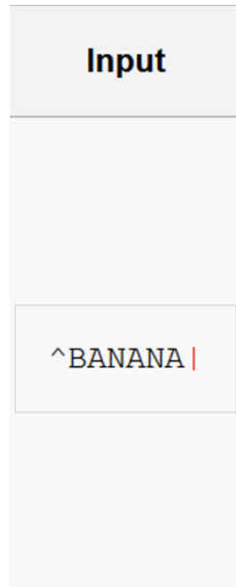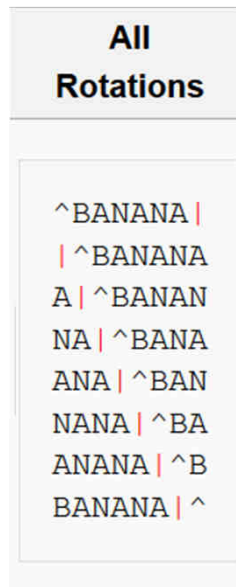
Burroughs-Wheeler transform indexing

Sorting All Rows in Alphabetical Order

```
ANANA|^B
ANA|^BAN
A|^BANAN
BANANA|^
NANA|^BA
NA|^BANA
^BANANA|
|^BANANA
```

Burroughs-Wheeler transform indexing

Taking Last Column

```
ANANA|^B
ANA|^BAN
A|^BANAN
BANANA|^
NANA|^BA
NA|^BANA
^BANANA|
|^BANANA
```

Burroughs-Wheeler transform indexing

Output **Last Column**	
BNN^AA|A	

Burroughs-Wheeler transform indexing

	Transformation			
Input	**All Rotations**	**Sorting All Rows in Alphabetical Order**	**Taking Last Column**	**Output Last Column**
^BANANA|	^BANANA| |^BANANA A|^BANAN NA|^BANA ANA|^BAN NANA|^BA ANANA|^B BANANA|^	ANANA|^B ANA|^BAN A|^BANAN BANANA|^ NANA|^BA NA|^BANA ^BANANA| |^BANANA	ANANA|^B ANA|^BAN A|^BANAN BANANA|^ NANA|^BA NA|^BANA ^BANANA| |^BANANA	BNN^AA|A

BWT is remarkable because it is
reversible.

Any ideas as how you might reverse it?

Burroughs-Wheeler transform indexing

Input
BNN^AA|A

Burroughs-Wheeler transform indexing

Add 1	Sort 1	Add 2	Sort 2
B	A	BA	AN
N	A	NA	AN
N	A	NA	A|
^	B	^B	BA
A	N	AN	NA
A	N	AN	NA
		^	|^	^B
A			A|	|^

Write the sequence as the last column · **Sort it…** · **Add the columns…** · **Sort those…**

Burroughs-Wheeler transform indexing

Add 3	Sort 3	Add 4	Sort 4
BAN	ANA	BANA	ANAN
NAN	ANA	NANA	ANA|
NA|	A|^	NA|^	A|^B
^BA	BAN	^BAN	BANA
ANA	NAN	ANAN	NANA
ANA	NA|	ANA|	NA|^
|^B	^BA	|^BA	^BAN
A|^	|^B	A|^B	|^BA

Add the columns… · **Sort those…** · **Add the columns…** · **Sort those…**

Burroughs-Wheeler transform indexing

Add 5	Sort 5	Add 6	Sort 6
BANAN	ANANA	BANANA	ANANA|
NANA|	ANA|^	NANA|^	ANA|^B
NA|^B	A|^BA	NA|^BA	A|^BAN
^BANA	BANAN	^BANAN	BANANA
ANANA	NANA|	ANANA|	NANA|^
ANA|^	NA|^B	ANA|^B	NA|^BA
|^BAN	^BANA	|^BANA	^BANAN
A|^BA	|^BAN	A|^BAN	|^BANA

Add the columns… · **Sort those…** · **Add the columns…** · **Sort those…**

http://en.wikipedia.org/wiki/Burrows-Wheeler_transform

Burroughs-Wheeler transform indexing

Add 7	Sort 7	Add 8
BANANA|	ANANA|^	BANANA|^
NANA|^B	ANA|^BA	NANA|^BA
NA|^BAN	A|^BANA	NA|^BANA
^BANANA	BANANA|	^BANANA|
ANANA|^	NANA|^B	ANANA|^B
ANA|^BA	NA|^BAN	ANA|^BAN
|^BANAN	^BANANA	|^BANANA
A|^BANA	|^BANAN	A|^BANAN

Add the columns… · **Sort those…** · **Add the columns…**

The row with the "end of file" character at the end is the original text

http://en.wikipedia.org/wiki/Burrows-Wheeler_transform

Burroughs-Wheeler transform indexing

Output
^BANANA|

**The row with the "end of file"
character at the end is the
original text**

http://en.wikipedia.org/wiki/Burrows-Wheeler_transform

Burroughs Wheeler indexing

Trapnell C, Salzberg SL, *Nat. Biotech.*, 2009