


Genome wiz Venter partners with Roche in DNA sequencing deal

BY JULIE STEENHUYSEN
CHICAGO | Wed Jan 14, 2015 3:35pm EST

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Another hat tossed into the million genome ring (so to speak)...

Human Longevity Inc (HLI), a start-up formed last March with the goal of sequencing 1 million genomes by 2020.

1 OF 2: Craig Venter, founder, chairman, and president of the J. Craig Venter Institute, listens to opening statements at House Energy and Commerce hearing on "Developments in Synthetic Genomics and Implications for Health and Energy" on Capitol Hill in Washington, in this file photo taken on May 27, 2010.
CREDIT: REUTERS/HYUNGWON KANG

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Reuters
1-14-2015

(Reuters) - Genome pioneer J. Craig Venter has signed a multi-year deal to sequence and analyze tens of thousands of genomes for Roche's Genentech unit in a deal aimed at identifying new drug targets and biomarkers, the companies said on Wednesday.

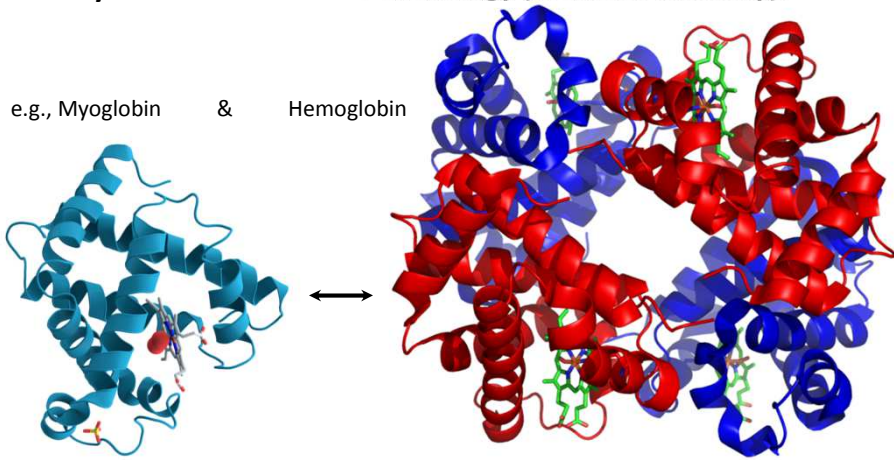
The deal is one of the biggest yet for Venter's La Jolla, California based Human Longevity Inc (HLI), a start-up formed last March with the goal of sequencing 1 million genomes by 2020. Financial terms were not disclosed.

Typically, to be “biologically related” means to share a common ancestor. In biology, we call this *homologous*.

Two proteins sharing a common ancestor are said to be *homologs*.

Homology often implies structural similarity & sometimes (not always) sequence similarity. **A statistically significant sequence or structural similarity can be used to infer homology (common ancestry).**

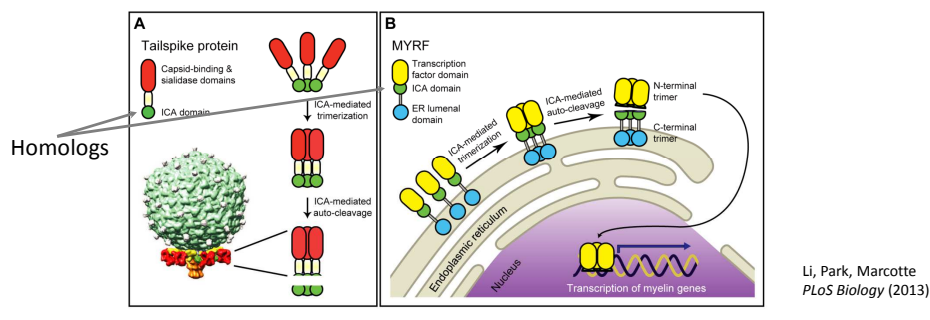
e.g., Myoglobin & Hemoglobin



<http://en.wikipedia.org/wiki/File:Myoglobin.png> & [File:1GZX_Haemoglobin.png](http://en.wikipedia.org/wiki/File:1GZX_Haemoglobin.png)

In practice, searching for sequence or structural similarity is one of the most powerful computational approaches for discovering functions for genes, since we can often glean many new insights about a protein based on what is known about its homologs.

Here's an example from my own lab, where we discovered that myelinating the neurons in your brain employs the same biochemical mechanism used by bacteriophages to make capsids. The critical breakthrough was recognizing that the human and phage proteins contained homologous domains.



Sequence alignment algorithms such as BLAST, PSI-BLAST, FASTA, and the Needleman–Wunsch & Smith-Waterman algorithms arguably comprise some of the most important driver technologies of modern biology and underlie the sequencing revolution.

So, let's start learning bioinformatics algorithms by learning how to align two protein sequences.

Live demo:

[http://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&BLAST_PROGRAMS=blastp
&PAGE_TYPE=BlastSearch&SHOW_DEFAULTS=on&LINK_LOC=blasthome](http://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&BLAST_PROGRAMS=blastp&PAGE_TYPE=BlastSearch&SHOW_DEFAULTS=on&LINK_LOC=blasthome)

MVLSPADKTNVKAAWGKVGAGHAGEYGAEALERMFSLFPTTKTYFPHFDLSHGSAQVKGHG
KKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLPAEFTP
AVHASLDDKFLASVSTVLTSKYR

Title: All non-redundant GenBank CDS translations+PDB+SwissProt+PIR+PRF excluding
environmental samples from WGS projects

Molecule Type: Protein

Update date: 2015/01/18

Number of sequences: 57851050

Protein sequence alignment

Two biologically related proteins with similar sequences:

FlgA1 EAGNVKLKRGRLDTLPPRTVLDINQLVDAISLRDLSPDQPIQLTQFRQAWRVKAGQQRVNVIASGD
++K+K+GRLDTLPP +L+ N A+SLR ++ QP+ R+ W +KAGQ V V+A G+
FlgA2 TLQDIKMKQGRDLTLPPGALLEPNFAQGAVSLRQINAGQPLTRNMLRRLWIIKAGQDVQVLALGE

Also biologically related (& fold up into the same 3D protein structure):

FlgA1 EAGNVKLKRGRLDTLPPRTVLDINQLVDAISLRDLSPDQPIQLTQFRQAWRVKAGQQRVNVIASGD
A + P +L I+ R L P + I R+AW V+ G V V
FlgA3 LAALKQVTTLIAGKHKPDAMATHAEELQGKIAKRTLLPGRYIPTAAIREAWLVEQGAQVFFFIAG

But these are biologically unrelated (& fold up into unrelated structures):

FlgA1 AGNVKLKRGRLDTLPPRTVLDINQLVDAISLRDLSPDQPIQLTQFRQA-WRVKAGQQRVNVIASGD
AG+V K G + + PRT ++ I+ P PI +++A WRV A + V V+ GD
HvcPP AGHV--KNGTMRIVGPRTCSNVWNGTFPINATTGPSIPI PAPNYKKALWRVSATEYVEVVRVGD

(FYI, we'll draw examples from Durbin *et al.*, *Biological Sequence Analysis*, Ch. 1 & 2).

To align two sequences, we need to perform 3 steps:

- 1. We need some way to decide which alignments are better than others.
For this, we'll invent a way to give the alignments a "score" indicating their quality.**
- 2. Align the two proteins so that they get the best possible score.**
- 3. Decide if the score is "good enough" for us to believe the alignment is biologically significant.**

To align two sequences, we need to perform 3 steps:

- 1. We need some way to decide which alignments are better than others.
For this, we'll invent a way to give the alignments a "score" indicating their quality.**
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We'll treat mutations as independent events.

This allows us to create an ***additive scoring scheme***.

The score for a sequence alignment will be the sum of the scores for aligning each of the individual positions in two sequences.

What kind of mutations should we expect?

Substitutions, insertions and deletions.

Insertions and deletions can be treated as equivalent events by considering one or the other sequence as the reference, and are usually called ***gaps***.

AGNVKLKRG
AG+V K G
AGHV - - KNG

substitution *gap*

Let's consider two models:

First, a **random** model, where amino acids in the sequences occur independently at some given frequencies.

The probability of observing an alignment between x and y is just the product of the frequencies (q) with which we find each amino acid.

We can write this as:

$$P(x, y | R) = \prod_i q_{x_i} \prod_j q_{y_j}$$

What's this mean? (pointing to $P(x, y | R)$)

What does the capital pi mean? (pointing to \prod)

What's this mean? (pointing to q_{x_i})

What's this mean? (pointing to q_{y_j})

Second, a **match** model, where amino acids at a given position in the alignment arise from some common ancestor with a probability given by the joint probability p_{ab} .

So, under this model, the probability of the alignment is the product of the probabilities of seeing the individual amino acids aligned.

We can write that as:

$$P(x, y | M) = \prod_i p_{x_i, y_i}$$

What's this mean? (pointing to $P(x, y | M)$)

What does the capital pi mean again? (pointing to \prod)

What's this mean? (pointing to p_{x_i, y_i})

To decide which model better describes an alignment, we'll take the ratio:

$$\frac{P(x, y | M)}{P(x, y | R)} = \frac{\prod_i P_{x_i y_i}}{\prod_i q_{x_i} \prod_j q_{y_j}} = \prod_i \frac{P_{x_i y_i}}{q_{x_i} q_{y_i}}$$

What did these mean again?

Such a ratio of probabilities under 2 different models is called an **odds ratio**.

Where else have you heard odds ratios used?

Basically: if the ratio > 1, model *M* is more probable
if < 1, model *R* is more probable.

Now, to convert this to an additive score *S*, we can simply take the logarithm of the odds ratio (called the **log odds ratio**):

$$S = \sum_i s(x_i, y_i)$$

This is just the score for aligning one amino acid with another amino acid:

$$s(a, b) = \log \left(\frac{P_{ab}}{p_a p_b} \right)$$

Here written *a* and *b* rather than *x_i* and *y_i* to emphasize that this score reflects the inherent preference of the two amino acids (*a* and *b*) to be aligned.

Almost done with step 1...

The last trick:

Take a big set of pre-aligned protein sequence alignments (that are correct!) and measure all of the pairwise amino acid substitution scores (the $s(a,b)$'s). Put them in a 20x20 **amino acid substitution matrix** :

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	5	-2	-1	-2	-1	-1	-1	0	-2	-1	-2	-1	-1	-3	-1	1	0	-3	-2	0
R	-2	7	-1	-2	-4	1	0	-3	0	-4	-3	3	-2	-3	-3	-1	-1	-3	-1	-3
N	-1	-1	7	2	-2	0	0	0	1	-3	-4	0	-2	-4	-2	1	0	-4	-2	-3
D	-2	-2	2	8	-4	0	2	-1	-1	-4	-4	-1	-4	-5	-1	0	-1	-5	-3	-4
C	-1	-4	-2	-4	13	-3	-3	-3	-3	-2	-2	-3	-2	-2	-4	-1	-1	-5	-3	-1
Q	-1	1	0	0	-3	7	2	-2	1	-3	-2	2	0	-4	-1	0	-1	-1	-1	-3
E	-1	0	0	2	-3	2	6	-3	0	-4	-3	1	-2	-3	-1	-1	-1	-3	-2	-3
G	0	-3	0	-1	-3	-2	-3	8	-2	-4	-4	-2	-3	-4	-2	0	-2	-3	-3	-4
H	-2	0	1	-1	-3	1	0	-2	10	-4	-3	0	-1	-1	-2	-1	-2	-3	2	-4
I	-1	-4	-3	-4	-2	-3	-4	-4	-4	5	2	-3	2	0	-3	-3	-1	-3	-1	4
L	-2	-3	-4	-4	-2	-2	-3	-4	-3	2	5	-3	3	1	-4	-3	-1	-2	-1	1
K	-1	3	0	-1	-3	2	1	-2	0	-3	-3	6	-2	-4	-1	0	-1	-3	-2	-3
M	-1	-2	-2	-4	-2	0	-2	-3	-1	2	3	-2	7	0	-3	-2	-1	-1	0	1
F	-3	-3	-4	-5	-2	-4	-3	-4	-1	0	1	-4	0	8	-4	-3	-2	1	4	-1
P	-1	-3	-2	-1	-4	-1	-1	-2	-2	-3	-4	-1	-3	-4	10	-1	-1	-4	-3	-3
S	1	-1	1	0	-1	0	-1	0	-1	-3	-3	0	-2	-3	-1	5	2	-4	-2	-2
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	2	5	-3	-2	0
W	-3	-3	-4	-5	-5	-1	-3	-3	-3	-3	-2	-3	-1	1	-4	-4	-3	15	2	-3
Y	-2	-1	-2	-3	-3	-1	-2	-3	2	-1	-1	-2	0	4	-3	-2	-2	2	8	-1
V	0	-3	-3	-4	-1	-3	-3	-4	-4	4	1	-3	1	-1	-3	-2	0	-3	-1	5

This is the **BLOSUM50** matrix.

(The numbers are scaled & rounded off to the nearest integer):

What's the score for aspartate (D) aligning with itself?

How about aspartate with phenylalanine (F)? Why?

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	5	-2	-1	-2	-1	-1	-1	0	-2	-1	-2	-1	-1	-3	-1	1	0	-3	-2	0
R	-2	7	-1	-2	-4	1	0	-3	0	-4	-3	3	-2	-3	-3	-1	-1	-3	-1	-3
N	-1	-1	7	2	-2	0	0	0	1	-3	-4	0	-2	-4	-2	1	0	-4	-2	-3
D	-2	-2	2	8	-4	0	2	-1	-1	-4	-4	-1	-4	-5	-1	0	-1	-5	-3	-4
C	-1	-4	-2	-4	13	-3	-3	-3	-3	-2	-2	-3	-2	-2	-4	-1	-1	-5	-3	-1
Q	-1	1	0	0	-3	7	2	-2	1	-3	-2	2	0	-4	-1	0	-1	-1	-1	-3
E	-1	0	0	2	-3	2	6	-3	0	-4	-3	1	-2	-3	-1	-1	-1	-3	-2	-3
G	0	-3	0	-1	-3	-2	-3	8	-2	-4	-4	-2	-3	-4	-2	0	-2	-3	-3	-4
H	-2	0	1	-1	-3	1	0	-2	10	-4	-3	0	-1	-1	-2	-1	-2	-3	2	-4
I	-1	-4	-3	-4	-2	-3	-4	-4	-4	5	2	-3	2	0	-3	-3	-1	-3	-1	4
L	-2	-3	-4	-4	-2	-2	-3	-4	-3	2	5	-3	3	1	-4	-3	-1	-2	-1	1
K	-1	3	0	-1	-3	2	1	-2	0	-3	-3	6	-2	-4	-1	0	-1	-3	-2	-3
M	-1	-2	-2	-4	-2	0	-2	-3	-1	2	3	-2	7	0	-3	-2	-1	-1	0	1
F	-3	-3	-4	-5	-2	-4	-3	-4	-1	0	1	-4	0	8	-4	-3	-2	1	4	-1
P	-1	-3	-2	-1	-4	-1	-1	-2	-2	-3	-4	-1	-3	-4	10	-1	-1	-4	-3	-3
S	1	-1	1	0	-1	0	-1	0	-1	-3	-3	0	-2	-3	-1	5	2	-4	-2	-2
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	2	5	-3	-2	0
W	-3	-3	-4	-5	-5	-1	-3	-3	-3	-3	-2	-3	-1	1	-4	-4	-3	15	2	-3
Y	-2	-1	-2	-3	-3	-1	-2	-3	2	-1	-1	-2	0	4	-3	-2	-2	2	8	-1
V	0	-3	-3	-4	-1	-3	-3	-4	-4	4	1	-3	1	-1	-3	-2	0	-3	-1	5

Using this matrix, **we can score any alignment as the sum of scores of individual pairs of amino acids.**

For example, the top alignment in our earlier example:

```

FlgA1 EAGNVKLKRGRD1TLPPRTVLDINQLVDAISLRDLSPDQPIQLTQFRQAWRVKAGQRVNVIASGD
      ++K+K+GRD2TLPP  +L+ N   A+SLR ++  QP+      R+ W +KAGQ V V+A G+
FlgA2 TLQDIKMKQGRD1TLPPGALLEPNFAQGAVSLRQINAGQPLTRNMLRRLWIIKAGQDVQVLALGE
  
```

gets the score:

$$S(\text{FlgA1}, \text{FlgA2}) = -1 - 2 - 2 + 2 + 4 + 6 + \dots = 186$$

We also need to penalize **gaps**. For now, let's just use a constant penalty ***d*** for each amino acid gap in an alignment, *i. e.*:

the penalty for a gap of length $g = -g*d$

PAM



Margaret Dayhoff (1925-1983)
Developed point accepted
mutation matrices
(PAM matrices)

Calibrated for different evolutionary times
PAM- n = n substitutions per 100 residues
e.g. matrices from PAM1 to PAM250
measure PAM1,
calculate higher PAMs from that

Explicit model of evolution
(calculated using a phylogenetic tree)

vs.

BLOSUM



Steve and Jorja Henikoff
Developed BLOSUM matrices

Calibrated for different % identity sequences
BLOSUM- n = for sequences of about n % identity
averages substitution probabilities over
sequence clusters, gives better estimates
for highly divergent cases

Implicit model of evolution
(calculated from blocks of aligned sequences)

To align two sequences, we need to perform 3 steps:

1. We need some way to decide which alignments are better than others.
For this, we'll invent a way to give the alignments a "score" indicating their quality.
2. Align the two proteins so that they get the best possible score.
3. Decide if the score is "good enough" for us to believe the alignment is biologically significant.

We'll use something called *dynamic programming*.

This is **mathematically guaranteed** to find the best scoring alignment, and uses *recursion*. This means problems are broken into sub-problems, which are in turn broken into sub-problems, etc, until the simplest sub-problems can be solved.

We're going to find the best **local** alignment—the best matching internal alignment—without forcing all of the amino acids to align (i.e. to match **globally**).

i.e., this \longrightarrow

ATGCAT
ATGCAT

Not this \longrightarrow

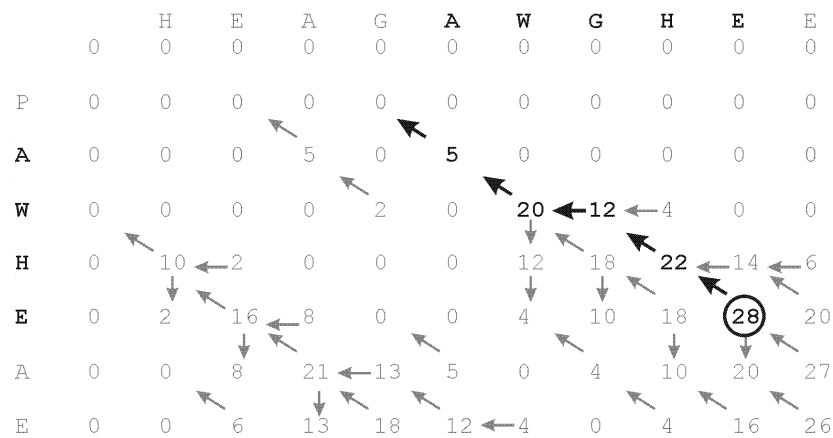
ACGTTATGCATGACGTA
-C---ATGCAT-----T-

Just two more rules:

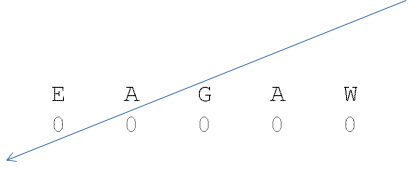
If the score is negative, set it equal to zero.

At each step, we also keep track of which event was chosen by **drawing an arrow from the cell we just filled back to the cell which contributed its score to this one.**

That's it! Just repeat this to fill the entire matrix.



Here we go! Start with the borders & the first entry.



		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0									
A	0										
W	0										
H	0										
E	0										
A	0										
E	0										

Why is this zero?

What's the score from our BLOSSUM matrix for substituting H for P?

Next round!

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0								
A	0	0	0								
W	0										
H	0										
E	0										
A	0										
E	0										

Terrible! Again, none of the possible give positive scores.

We have to go a bit further in before we find a positive score...

A few more rounds, and a positive score at last!

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0							
A	0	0	0	5							
W	0	0	0								
H	0										
E	0										
A	0										
E	0										

How did we get this one?

& a few more rounds...

		H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0						
A	0	0	0	5	0						
W	0	0	0	0	2						
H	0	10	2	0	0						
E	0										
A	0										
E	0										

What does this mean?

The whole thing filled in!

	0	H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	5	0	5	0	0	0	0	0
W	0	0	0	0	2	0	20	12	4	0	0
H	0	10	2	0	0	0	12	18	22	14	6
E	0	2	16	8	0	0	4	10	18	28	20
A	0	0	8	21	13	5	0	4	10	20	27
E	0	0	6	13	18	12	4	0	4	16	26

Now, find the optimal alignment using a **traceback** process:

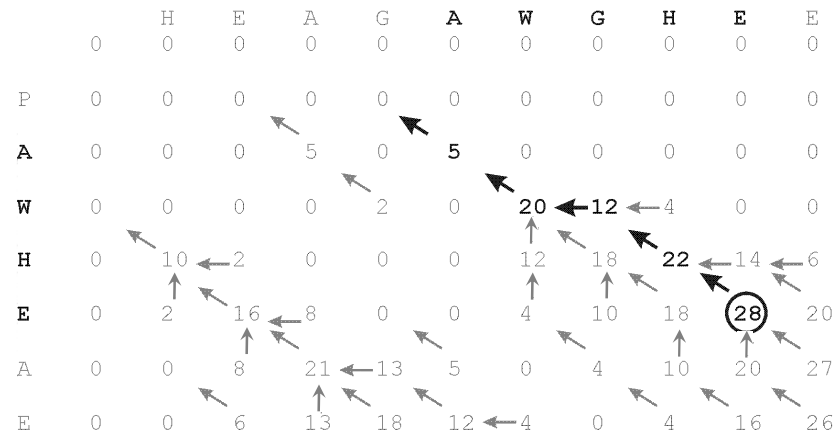
Look for the highest score, then follow the arrows back.

The alignment “grows” from right to left

	0	H	E	A	G	A	W	G	H	E	E
	0	0	0	0	0	0	0	0	0	0	0
P	0	0	0	0	0	0	0	0	0	0	0
A	0	0	0	5	0	5	0	0	0	0	0
W	0	0	0	0	2	0	20	12	4	0	0
H	0	10	2	0	0	0	12	18	22	14	6
E	0	2	16	8	0	0	4	10	18	28	20
A	0	0	8	21	13	5	0	4	10	20	27
E	0	0	6	13	18	12	4	0	4	16	26

This gives the following alignment: A W G H E
 A W - H E

(Note: for gaps, the arrow points to the sequence that gets the gap)



To align two sequences, we need to perform 3 steps:

1. We need some way to decide which alignments are better than others.
 For this, we'll invent a way to give the alignments a "score" indicating their quality.
2. Align the two proteins so that they get the best possible score.
3. Decide if the score is "good enough" for us to believe the alignment is biologically significant.

This algorithm always gives the best alignment.

Every pair of sequences can be aligned in some fashion.

So, when is a score “good enough”?

How can we figure this out?

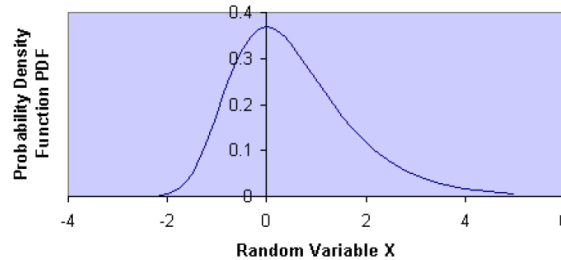
Here's one approach:

**Shuffle one sequence. Calculate the best alignment & its score.
Repeat 1000 times.**

If we never see a score as high as the real one, we say the real score has < 1 in a 1000 chance of happening just by luck.

But if we want something that only occurs < 1 in a million, we'd have to shuffle 1,000,000 times...

Luckily, alignment scores follow a well-behaved distribution, the **extreme value distribution**, so we can do a few trials & fit to this.



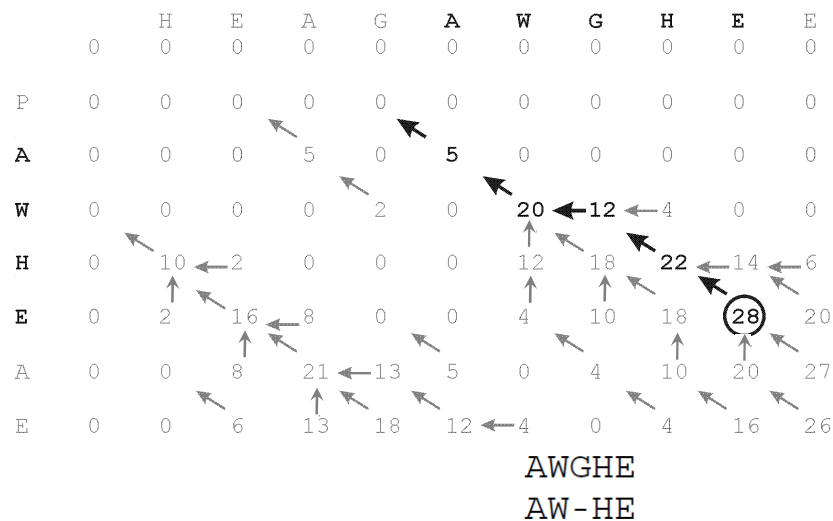
random trials & their average score

$$p(\text{max score} \leq X) \approx e^{-kNe^{\lambda(X-\mu)}}$$

This p-value gives the significance of your alignment.
But, if we search a database and perform many alignments, we still need something more (next time).

Describe the shape & can be fit from a few trials

Some extensions: Local vs. global alignments
How might you force the full sequences to align?



Some extensions: Local vs. global alignments
How might you force the full sequences to align?

A few tiny changes:

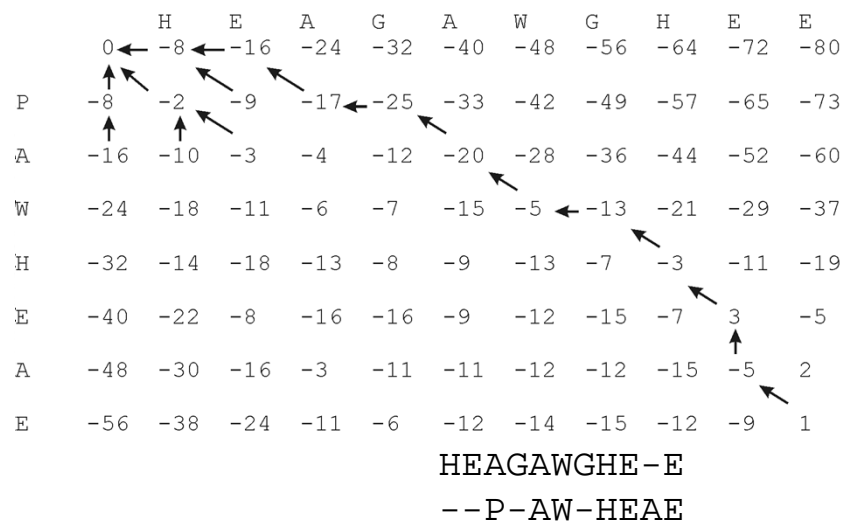
Initialize only the top left cell of the path matrix to zero
(not all top and left cells).

Leave the negative values (don't set them to zero).

The optimal alignment should start at the top left cell and
finish at the bottom right cell of the path matrix.

Start the trace-back at the bottom right cell

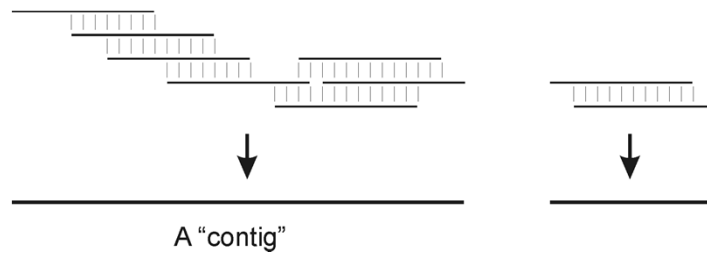
Some extensions: Local vs. global alignments
How might you force the full sequences to align?



Some extensions:

What about overlapping sequences?

e.g. as in 'shotgun sequencing' genomes where
'contigs' are built up from overlapping sequences



Some extensions:

What about overlapping sequences?

Modify global alignment to not penalize overhangs:

The optimal alignment should start at the top or left edge
and finish at the bottom or right edge of the path matrix.

Set these boundary conditions :

$$F(i, 0) = 0 \text{ for } i=1 \text{ to } n$$

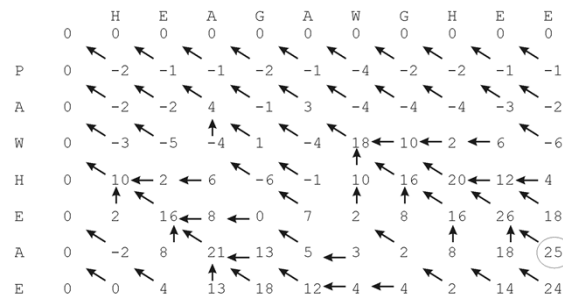
$$F(0, j) = 0 \text{ for } j=1 \text{ to } m$$

Start the traceback at the cell with the highest score on the
right or bottom border

Some extensions:

What about overlapping sequences?

e.g. as in 'shotgun sequencing' genomes where
'contigs' are built up from overlapping sequences



(overhang = HEA)

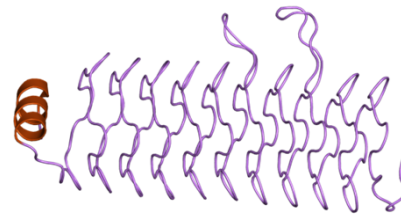
GAWGHEE

PAW-HEA

(overhang = E)

Some extensions:

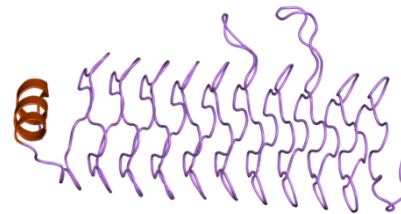
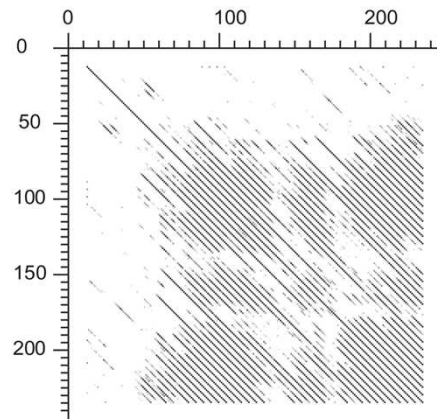
How might you find repetitive sequences?



Structure of the pentapeptide
repeat protein HetL

(from wiki, PMID18952182)

Align the sequence to itself and ignore the diagonal (optimal) alignment
→ High-scoring off-diagonal alignments will be repeats



Structure of the pentapeptide
repeat protein HetL
(from wiki, PMID18952182)

Dot plot (quick visualization of
sequence similarity)
of the pentapeptide repeat
protein HgIK protein vs. itself
(http://en.wikipedia.org/wiki/Pentapeptide_repeat)